

Serum and Tissue Zinc in Epithelial Malignancies: A Meta-Analysis



Jaromir Gumulec^{1,2}, Michal Masarik^{1,2*}, Vojtech Adam^{2,3}, Tomas Eckschlager⁴, Ivo Provaznik⁵, Rene Kizek^{2,3}

1 Department of Pathological Physiology, Faculty of Medicine, Masaryk University, Brno, Czech Republic, 2 Central European Institute of Technology, Brno University of Technology, Brno, Czech Republic, 3 Department of Chemistry and Biochemistry, Mendel University in Brno, Brno, Czech Republic, 4 Department of Paediatric Haematology and Oncology, 2nd Faculty of Medicine and University Hospital Motol, Charles University, Prague, Czech Republic, 5 Department of Biomedical Engineering, Faculty of Electrical Engineering and Communication, Brno University of Technology, Brno, Czech Republic

Abstract

Background and Objectives: Current studies give us inconsistent results regarding the association of neoplasms and zinc(II) serum and tissues concentrations. The results of to-date studies using meta-analysis are summarized in this paper.

Methods: Web of Science (Science citation index expanded), PubMed (Medline), Embase and CENTRAL were searched. Articles were reviewed by two evaluators; quality was assessed by Newcastle-Ottawa scale; meta-analysis was performed including meta-regression and publication bias analysis.

Results: Analysis was performed on 114 case control, cohort and cross-sectional studies of 22737 participants. Decreased serum zinc level was found in patients with lung (effect size = -1.04), head and neck (effect size = -1.43), breast (effect size = -0.93), liver (effect size = -0.93), liver (effect size = -0.93), stomach (effect size = -0.93), and prostate (effect size = -0.93), and proven in any tumor. More specific zinc patterns are evident at tissue level, showing increase in breast cancer tissue (effect size = -0.93) and decrease in prostatic (effect size = -0.93), liver (effect size = -0.93), lung (effect size = -0.93), and thyroid cancer (effect size = -0.93). The rest of the included tumors brought ambiguous results, both in serum and tissue zinc levels across the studies. The association between zinc level and stage or grade of tumor has not been revealed by meta-regression.

Conclusion: This study provides evidence on cancer-specific tissue zinc level alteration. Although serum zinc decrease was associated with most tumors mentioned herein, further – prospective - studies are needed.

Citation: Gumulec J, Masarik M, Adam V, Eckschlager T, Provaznik I, et al. (2014) Serum and Tissue Zinc in Epithelial Malignancies: A Meta-Analysis. PLoS ONE 9(6): e99790. doi:10.1371/journal.pone.0099790

Editor: Marcia E Lopes. Consolaro, State University of Maringá/Universidade Estadual de Maringá, Brazil

Received February 6, 2014; Accepted May 18, 2014; Published June 18, 2014

Copyright: © 2014 Gumulec et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: Financial support from Central European Institute of Technology CZ.1.05/1.1.00/02.0068 and Internal Grant Agency of Ministry of Health of the Czech Republic (IGA MH NT14337-3/2013) is highly acknowledged. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

1

Competing Interests: The authors have declared that no competing interests exist.

* Email: masarik@med.muni.cz

Introduction

Zinc(II) plays a role in several intracellular signalling pathways. It is also a cofactor of numerous enzymes [1]. Its dysregulation is present in various cancers. Imbalance of zinc transporters causing intracellular and serum zinc(II) levels alteration was described in prostate and breast cancers. [2–5]. Questions were raised whether these associations have clinical applications. Studies focusing on zinc content in biological materials in cancer patients provide inconsistent results. Zinc levels in tumor tissues of prostate [6], liver [7], and lung [8] and its serum levels in breast, lung, stomach, and prostate cancer patients were reviewed previously [9].

We investigated the associations of serum and cellular zinc(II) levels with carcinomas via meta-analysis.

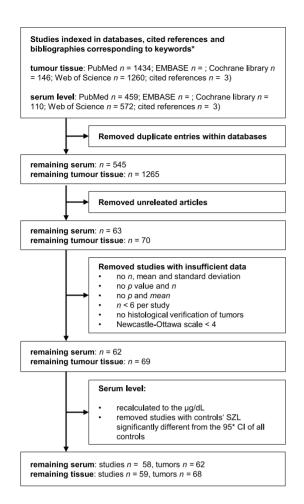
Methods

Literature Search

Search was performed in Web of science (Science citation index expanded 1945 to April 2013), PubMed (Medline 1968 to April 2013), Embase (1977 to April 2013), and Cochrane Library (CENTRAL 1953 to April 2013); keywords are shown in Figure 1. Moreover, cited references of found articles were analyzed.

Selection Criteria

Diagram (Fig. 1) shows acquisition process. Among published articles, the search was done for clinical trials, case-control and cohort studies investigating the associations between carcinoma and tissues and serum zinc levels. Because no difference in zinc(II) level between serum and plasma was found [10], both materials were referred as "serum". Studies with full texts available were



*keywords:

(zinc OR zn) serum (cancer OR tumour OR tumor OR neoplas*) + (zinc OR zn) tissue cancer (cancer OR tumour OR tumor OR neoplas*) +

bladder/Breast/Cervi*/Colorect*/Colon/Rectum/Esophag*/Gallbladder/ Head Neck/Gynecolog*/I/Kidney/Laryn*/ Lung//Pancrea*/Pharyn*/Prostat*/Salivary gland/Stomach/Thyroid*/Uter*

Figure 1. Flow diagram for identification of relevant studies. doi:10.1371/journal.pone.0099790.q001

included only. Only the studies where the data were displayed in the following ways were accepted: (1) sample size, means and standard deviations, or (2) sample size, means, P values and statistical test type (one- or two-tailed). If similar data were found in more studies by the same group, study with most data was included.

The eligibility of the studies for meta-analysis was evaluated by J.G. and V.A., discrepancies were discussed with R.K and M.M.

Assessment of Methodological Quality

The quality of studies was assessed by the Newcastle-Ottawa Scale (NOS) [11]. NOS ranges from 0 to 9 stars. The studies with <4 stars were excluded, with >6 stars were considered as high quality; the mean was 5.6 stars.

Main and Subgroup Analyses

First, differences of the serum and tissue zinc levels between overall tumors and controls were analysed (Table 1). Then, analyses by tumor type, histology, and methodological quality were performed (Table 2). To meet conditions of between-study

independence, zinc level was averaged in studies with multiple tumors, forming age groups or detecting gender separately in case of the summary. For comparison of individual tumors, the tumor was taken as a unit of analysis unless violated between-study independence.

Statistical Analyses

To express the differences in serum and tissue concentrations, standardized difference in means (Cohen's d) was used. To assess heterogeneity across studies, Higgins I² (describes percentage of variability) was calculated [12]. Random effects model meta-analysis was employed when I²>50%; otherwise, fixed model was used. Publication bias was evaluated using funnel plots and two-sided Egger tests in groups with >3 studies. Funnel plots of studies with Egger's test p<0.05 are asymmetric (Table S1). There was performed meta-regression using unrestricted maximum likehood method of studies reporting stage or grade if number of studies with corresponding moderators was >10 [13,14]. Comprehensive Meta-analysis Version 2 software (Biostat, Englewood, NJ) was used for analysis.

Results

Identification of Relevant Studies and their Characteristics

Total of 3201 articles were found. After excluding articles not meeting the criteria (Fig. 1) and duplicates, 114 articles studying 130 tumors were included (several studies studied more tumors, Table S2).

Overall 114 studies including 22737 participants (8584 cases, 14153 controls) were analyzed. From studies reporting age, sex and ethnicity, the mean age was 54.5 ± 12.3 (male) and 49.6 ± 12.2 (female); male were included 49.5% and 51.0% in "tumor" and control groups. Publication date ranged within 1952-2012. Caucasians, Asians, Hispanics, and Afro-Americans were reported in 52%, 45%, 1%, and 2%, respectively. Characteristics of studies are summarized in Table S2. Two clinical, 4 cohort, 6 cross-sectional, and 102 case control studies were included.

Overall Zinc Level in Sera and Tumors

Serum level. As shown on Fig. 2 and Table 1, serum zinc level is significantly decreased in patients with tumors (effect size = -1.08; 95% confidence interval, CI, -1.33 to -0.82) using random effects model of 58 studies (6223 cases, 10364 controls). This is consistent with subgroup of 26 high-quality studies (effect size = -1.30; 95% CI, -1.72 to -0.88). High level of heterogeneity is observed (Higgins $I^2 = 96.71$ %). Meta-regression did not reveal that stage, grade, and age or publication year affect effect size (Table S1). Six studies analyzed serum zinc level in group of patients with malignant tumor without other specification (516 cases, 3871 controls), and significant decrease was found (Fig. 2) [15–20].

Tissue level. There was a significant decrease in tissue zinc level using random effects model meta-analysis of 59 studies (2361 cases, 3789 controls) with effect size -1.44 (CI-1.93 to -0.95). However, publication bias was observed at p=0.01 and no significance found in 24 high-quality studies. Meta-regression did not reveal any moderators to affect global effect size.

Bladder

Significant decrease of serum zinc level was observed (-1.24; 95% CI, -1.77 to -0.77) using random effects model of two studies [21,22] (86 cases, 92 controls); both were "high-quality".

Table 1. Overall results of meta-analysis by tumor type and statistical model used.

Tumor	No. of studies Point estimate		95% CI of point estimate	Heterogeneity, I ²	Model used	
Bladder - serum	2	-1.24	-1.77 to -0.71	61.74	random	
Breast serum	12	-0.93	−1.68 to −0.17	96.16	random	
Breast tissue	15	1.80	1.17 to 2.42	94.89	random	
Colorectal serum	5	0.04	-2.57 to 2.64	98.98	random	
Colorectal tissue	7	0.37	-0.97 to 1.72	96.23	random	
Esophageal serum	4	-2.17	−3.23 to −1.11	86.46	random	
Esophageal tissue	3	-1.57	-3.17 to 0.03	94.72	random	
Gallbladder - serum	1	-2.31	−2.96 to −1.65		-	
Gallbladder - tissue	2	-1.25	−1.73 to −0.77	0.00	fixed	
Gynecological serum	3	-0.39	−0.6 to −0.17	49.48	fixed	
Gynecological tissue	4	-0.70	-1.85 to 0.45	91.40	random	
Head and neck tissue	2	3.11	-5.96 to 12.17	98.75	random	
Head Neck serum	5	-1.43	−2.17 to −0.68	77.46	random	
Kidney - tissue	4	-2.23	−3.89 to −0.57	89.51	random	
Liver - serum	3	-2.29	-5.21 to 0.63	97.64	random	
Liver - tissue	7	-8.26	−11.02 to −5.49	98.49	random	
Lung - serum	13	-1.04	−1.53 to −0.56	92.94	random	
Lung - tissue	6	-3.12	−4.57 to −1.67	96.76	random	
Prostate serum	7	-1.36	−1.97 to −0.75	97.93	random	
Prostate tissue	12	-3.90	−5.26 to −2.54	94.67	random	
Stomach serum	4	-1.59	−3.14 to −0.03	98.24	random	
Stomach tissue	3	-0.79	-1.44 to -0.14	60.85	random	
Thyroid serum	3	-0.62	-3.04 to 1.79	97.69	random	
Thyroid tissue	3	-2.84	−5.39 to −0.29	97.56	random	
Overall - serum	58	-1.08	-1.33 to -0.82	96.71	random	
Overall - tissue	59	-1.44	−1.93 to −0.95	97.08	random	

CI, confidence interval.

doi:10.1371/journal.pone.0099790.t001

Breast

Serum. There was significant decrease (-0.93; 95% CI, -1.68 to -0.17) using random effects model meta-analysis of 12 studies (604 cases, 663 controls) [20,23–33] (Fig. 3, Table 1). However, no significant change was observed in six studies [20,23,25,27,30,31] and significantly increased level in one [24]. No publication bias was observed (Begg's funnel plot was symmetrical, Egger's 2-tailed test p=0.086). Subgroup meta-analysis by methodological quality of study revealed significant decrease in nine low-quality studies (-1.32, CI-2.23 to-0.42) using random effects model, but no significant change in three high-quality studies using fixed effects model [25,27,31]. With regard to stage, Yucel *et al.* found no difference between stages [33]; in contrary, studies by Gupta *et al.* and Kuo *et al.* showed a decrease in advanced cancer in comparison to early stages [26] and significantly decreasing trend in relation to stage [29].

Tissue level. Significant elevation was determined using random effects model of 15 studies (635 cases, 714 controls), effect size = 1.80 (95% CI, 1.17 to 2.42) [29,34–47]. However, two studies show insignificant changes [42,46]. The highest levels were observed in study by Ng *et al.*, which include ductal cancers only [35]. Publication bias was observed on p = 0.018. Significant elevation was found in both low- and high- quality studies, while the levels were higher in 9 high-quality studies (2.33; CI, 1.46 to 3.21). Farquharson *et al.* reported significantly decreased tissue zinc

concentration in estrogen receptor negative tumors [42]. Kuo *et al.* found no trend [29] in relation to grade, while Farquharson *et al.* showed significant decrease in grade II-III vs. grade I [42].

Gynecological Tumors (Uterine Corpus, Cervix and Ovarian)

Serum. Among ovarian and cervical cancers, there was a significant decrease found (-0.39; CI-0.60 to -0.17) using fixed effects model of three studies (164 cases, 171 controls) [20,48,49]. No publication bias was observed. When subgroup meta-analysis by histological type was done, significant decrease was found in cervical [48], insignificant changes in ovarian cancer [49]. There were no significant trends in relation to stage and grade [49].

Tissue level. No significant difference between uterine corpus and cervix cancers was determined using random-effects model of 4 studies (80 cases, 123 controls) [37,48,50,51]. No publication bias was determined. Significant decrease was reported in two studies [48,51]. Results of high-quality studies did not show a significant trend.

Digestive System Tumors (Esophageal, Stomach, Colorectal, Liver, Gallbladder and Pancreatic Carcinoma)

Esophageal, serum. All four studies included in this analysis (93 cases, 80 controls) [52–55] show significant reduction (-2.17;

Table 2. Subgroup analysis by study quality and histological type.

Subgroup	Tumor	Factor	No. of studies	Point estimate (95% CI)	Heterogeneity, I ²	Model used
Study quality						
	Breast serum	high	3	0.22 (-0.08 to 0.51)	0.00	fixed
	Breast serum	low	9	-1.32 (-2.23 to -0.42)	97.06	random
	Breast tissue	high	9	2.33 (1.46 to 3.21)	95.88	random
	Breast tissue	low	6	1.11 (0.08 to 2.14)	91.22	random
	Colorectal tissue	high	2	-0.50 (-2.84 to 1.85)	81.73	random
	Colorectal tissue	low	5	0.80 (-0.84 to 2.43)	96.55	random
	Esophageal serum	high	2	-1.56 (-3.23 to 0.10)	72.04	random
	Esophageal serum	low	2	-2.99 (-4.79 to -1.19)	93.50	random
	Esophageal tissue	high	2	-1.60 (-3.86 to 0.65)	97.29	random
	Esophageal tissue	low	1	-1.52 (-2.56 to -0.47)		-
	Gynecological tissue	high	3	-0.14 (-1.02 to 0.75)	82.33	random
	Gynecological tissue	low	1	-2.49 (-3.31 to -1.66)		-
	Head Neck serum	high	2	−0.88 (−1.26 to −0.49)	3.16	fixed
	Head Neck serum	low	3	-1.59 (-2.08 to -1.10)	86.60	random
	Kidney - tissue	high	2	-1.03 (-1.80 to -0.26)	22.30	fixed
	Kidney - tissue	low	2	-3.12 (-6.49 to 0.25)	96.07	random
	Liver - tissue	high	2	-8.03 (-15.12 to -0.94)	98.00	random
	Liver - tissue	low	5	-12.29 (-17.08 to -7.51)	98.84	random
	Lung - serum	high	7	-1.01 (-1.69 to -0.33)	95.95	random
	Lung - serum	low	6	-1.09 (-1.84 to -0.34)	70.97	random
	Prostate serum	high	5	-2.07 (-3.82 to -0.32)	98.59	random
	Prostate serum	low	2	-0.34 (-3.07 to 2.39)	78.33	random
	Prostate tissue	high	3	-6.25 (-9.71 to -2.78)	98.02	random
	Prostate tissue	low	9	-3.59 (-5.34 to -1.85)	92.34	random
	Stomach serum	high	1	-0.13 (-0.32 to 0.06)	72.0	-
	Stomach serum	low	3	-2.09 (-4.14 to -0.04)	97.68	random
	Stomach tissue	high	2	-1.14 (-1.55 to -0.73)	0.00	fixed
	Stomach tissue	low	1	-0.26 (-0.96 to 0.45)	0.00	-
	Overall - serum	high	26	-1.30 (-1.72 to -0.88)	96.62	random
	Overall - serum	low	32	-0.92 (-1.3 to -0.54)	96.77	random
	Overall - tissue	high	23	-0.42 (-1.23 to 0.38)	97.38	random
			36			
distalasiani tus	Overall - tissue	low	30	-2.12 (-2.77 to -1.47)	96.95	random
Histological ty _l		0	1	0.07 (0.47 +- 0.22)		
	Gynecological serum	Ovary	1	-0.07 (-0.47 to 0.32)		-
	Gynecological serum	Uterine cervix	1	-0.49 (-0.77 to -0.21)	01.06	-
	Head Neck serum	Larynx	4	-1.52 (-2.35 to -0.69)	81.96	random
	Head Neck serum	Oral Cavity	2	-0.95 (-1.38 to -0.53)	0.00	fixed
	Lung - serum	adenocarcinoma 	2	0.02 (-1.49 to 1.54)	95.48	random
	Lung - serum	large cell	2	-0.78 (-1.32 to -0.23)	0.00	fixed
	Lung - serum	NSCLC 	4	-0.94 (-1.36 to -0.53)	58.87	random
	Lung - serum	small cell	2	-1.42 (-3.05 to 0.21)	88.46	random
	Lung - serum	squamous cell	2	-1.23 (-2.75 to 0.29)	97.01	random
	Lung - tissue	adenocarcinoma 	2	-0.89 (-2.96 to 1.19)	92.20	random
	Lung - tissue	large cell	1	-1.22 (-2.22 to -0.23)		-
	Lung - tissue	NSCLC	2	-0.62 (-1.88 to 0.65)	88.90	random
	Lung - tissue	small cell	2	-0.52 (-0.80 to -0.23)	0.00	fixed
	Lung - tissue	squamous cell	2	-0.48 (-2.54 to 1.58)	92.14	random
	Thyroid serum	folicular	2	-1.86 (-4.83 to 1.11)	96.81	random

Table 2. Cont.

Subgroup	Tumor	Factor	No. of studies	Point estimate (95% CI)	Heterogeneity, I ²	Model used
	Thyroid serum	papillary	2	-1.48 (-4.43 to 1.48)	96.38	random
	Thyroid tissue	folicular	1	0.00 (-0.59 to 0.59)		-
	Thyroid tissue	papillary	1	-0.50 (-1.09 to 0.09)		-

Note low is for Newcastle-Ottava scale <6, high for NOS >6. Random and fixed effect meta-analysis. NSCLC, non-small cell lung cancer, Cl, confidence interval. doi:10.1371/journal.pone.0099790.t002

95% CI, -3.23 to 1.11) and it was one of the highest decreases. Publication bias was identified (p = 0.04) and insignificant decrease was observed in two high-quality studies.

Esophageal, tissue. No significant change was identified using meta-analysis of three studies (104 cases, 116 controls) [52,56,57] due to high variability among studies, even among two high-quality ones [56,57]. No publication bias was observed.

Stomach, serum. Significant decrease was observed (effect size = -1.59; 95% CI, -3.14 to -0.03) without publication bias. However, high variability was present in the four studies (290 cases, 474 controls) [28,30,58,59] and significant decrease was reported in two studies only [28,59]. No significant change was observed in one high-quality study [58].

Stomach, tissue. Significant decrease was determined (-0.79; 95% CI, -1.44 to -0.14) in three studies (71 cases, 67 controls) [37,59,60] and one study showed significant decrease [59]. No publication bias was identified.

Colorectal cancer, serum. No significant difference was determined. No study fulfilled criteria of "high-quality". Five studies were included (313 cases, 216 controls) [28,30,61–63] and no publication bias was detected. However, two studies showed

significant decrease [28,62], one study showed significant elevation [61], and thus, high serum zinc level variances were associated with colorectal cancer. Of studies reporting stage, one showed significantly decreasing trend on Dukes stage [62], whereas other showed no significant changes related to TNM stage [61].

Colorectal cancer, tissue. No significant difference was observed using meta-analysis of seven studies (233 cases, 159 controls) [34,37,46,62,64–66] and no publication bias was observed. None of the high quality studies revealed significant differences. One of the largest variation among tissue levels was observed (effect size = 0.37; 95% CI, -0.97 to 1.72). Two studies showed significant decrease [62,64] while another two significant elevation [46,66]. One study including grade did not show significant trend [64].

Gallbladder, serum. one high-quality study (30 cases, 30 controls), that showed significant decrease, was identified [67].

Gallbladder, tissue. Significant decrease was determined in two studies [68,69] (39 cases, 40 controls) using fixed effects model (-1.25; 95% CI, -1.73 to -0.77). No study analyzed further classifications.

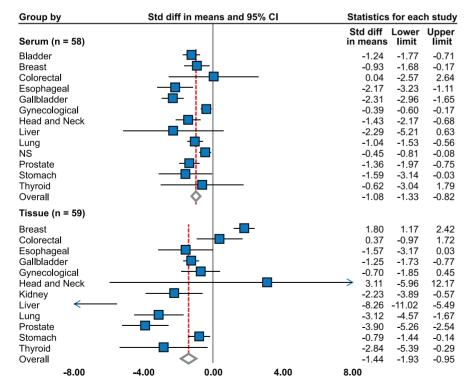


Figure 2. Level of zinc in sera and tissues by tumor type. Summary of individual meta-analyses. For model used and heterogeneity, see Table 1. doi:10.1371/journal.pone.0099790.g002

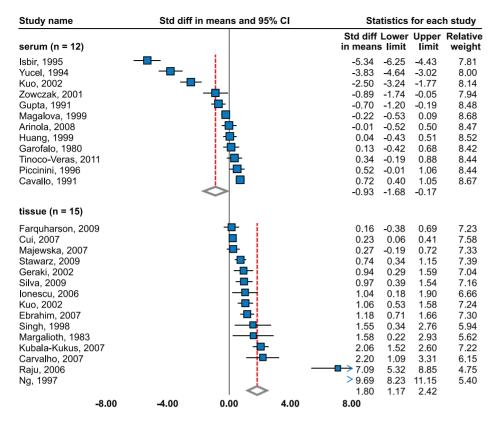


Figure 3. Zinc level in sera and tissues of breast cancer patients. Random effects model meta-analysis. Studies sorted by standardized mean difference.

doi:10.1371/journal.pone.0099790.g003

Liver, serum. No significant change was observed in three studies [70–72] (149 cases, 121 controls) and no publication bias was observed. Two high-quality studies [71,72] are in accordance, showing fixed serum zinc level. One study showed no significant trend in relation to stage [70].

Liver, tissue. Significant decrease was observed in all 7 studies analysed [70,73-78] (269 cases, 329 controls), effect size = -8.26 (95% CI, -11.02 to -5.49). Two high-quality studies [73,76] provided consistent results (-8.03; 95% CI, -15.12 to -0.93). Publication bias was observed (p = 0.04).

Prostate

Serum. Significant decrease was observed (effect size = -1.36; 95% CI, -1.97 to -0.75) using random effects model-meta-analysis of 7 studies [79–85] (2985 cases, 3539 controls, Fig. 4).

Insignificant changes were observed in large sample studies – cohort from French SuViMax study (4961 cases and controls) [82] and Multiethnic Cohort study (1175 cases and controls) [85]. Subgroup meta-analysis found significant decrease in high quality studies (-2.07; 95 CI, -3.82 to -0.32). No publication bias was observed

Tissue. Random effects model meta-analysis of 12 studies (240 cases, 226 controls) [86–97] detected a significant decrease (effect size = -3.90; 95% CI, -5.26 to -2.54). Only one study showed insignificant decrease [86]. Most significant decrease was observed in study by Guntupalli et al. [89]. High level of publication bias was observed among studies (p = 0.0007). Results are in agreement with more distinct effect size in high-quality studies (-6.03; 95% CI, -9.39 to -2.67). Trend was not observed in one study relating to stage and grade [6].

Head and Neck

Serum. Five studies including cancers of oral cavity [98,99] and larynx [98,100–102] (159 cases, 228 controls) showed significant decrease (random effects model -1.43; 95% CI, -2.17 to -0.68). No publication bias was observed - all studies, including 2 high-quality ones, showed significant decrease [98,99]. No significant trend was observed between stage and serum zinc level [100].

Tissue. Two studies (45 cases, 27 controls) [103,104] were included in the analysis. Findings of these studies were contradictory: one showed significant elevation [103] while the other significant decrease [104]. Random effects model did not show any trend. No significant trend between grade and zinc level was observed in one study [104].

Thyroid

Serum. No significant difference was observed using random effects model of three studies (131 cases, 93 controls) [105–107]. One study showed significant elevation [107], the other showed significant decrease [105] and third, ranked as high quality [106], found no significant differences. No publication bias was observed. Subgroup analysis by histological type does not highlight significant difference between papillary and follicular cancer. Medullar carcinoma was not included in meta-analysis.

Tissue. Statistically significant decrease was observed using random effects model (effect size = -2.84; 95% CI, -5.39 to -0.29) of three studies (109 cases, 123 controls) [105,108,109]. However, one study reported insignificant results [105], another included papillary, follicular, medullar cancers and reticulosarcoma [108] and the third did not specify histological types. No

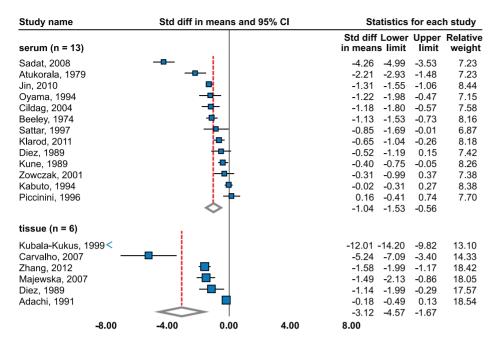


Figure 4. Zinc level in sera and tissues of prostate cancer patients. Random effects model meta-analysis. Studies sorted by standardized mean difference.

doi:10.1371/journal.pone.0099790.g004

publication bias was present. One study determined no significant difference between papillary and follicular cancers [105].

Kidney

Tissue. Significant decrease was observed using random effects model (-2.23; 95% CI, -3.89 to -0.57) of 4 studies (66 cases, 45 controls). Results agree with 2 high quality studies [37,110]. All studies showed significant decrease [37,110-112] and no publication bias was detected.

Lung

Serum. Significant decrease (-1.04; 95% CI, -1.53 to -1.530.56) was identified using random effect model of 13 studies [20,31,58,113–122] (703 cases, 786 controls, Fig. 5); four of them showed insignificant changes [20,31,58,115]. No publication bias was observed (p = 0.38). Analysis of only high-quality studies provided similar results. Subgroup analysis according to histology detected significant decrease in non-small cell lung cancer using random effects model in four studies. Two studies dealt with histological classification [115,116]: no significant difference was observed in adenocarcinoma and squamous cell carcinoma using random effects and significant decrease in large cell carcinoma using fixed effects (-0.78; 95% CI, -1.32 to -0.23). serum zinc level and stage was analyzed in two studies [114,116]. Klarod et al. determined significantly lower serum zinc level in advanced compared to low stages [114]. Similarly, descending trend was observed between stages T1, T2, and T3 [116]. Negative correlation between serum zinc level and grade was determined

Tissue. Significant decrease was determined using random effects model (-3.12; 95% CI, -4.57 to -1.67) of six studies, all ranked as low-quality [34,46,115,123-125] (470 cases, 1820 controls). However, publication bias was observed (p = 0.03). Insignificant change was observed in one study [123]. Significant decrease was determined in small cell lung cancer (-0.52; 95% CI, -0.80 to -0.23) using fixed model and no significant decrease was identified in non-small cell lung cancer using random effects

model. No significant trend was observed in squamous and adenocarcinomas. Large cell cancer showed significant decrease in one study [115].

Discussion

Decreased serum zinc level was found in patients with lung, head and neck, breast, liver, stomach, and prostate cancers. The elevation was not proven in any tumor. More specific zinc patterns are evident in tumors. Unequivocal increase was observed in breast cancer tissue only and decrease in prostatic, liver, lung, and thyroid cancer. The rest of the studied tumors brought ambiguous results, both in serum and tissue zinc levels across the studies. It cannot be confirmed that the serum zinc level does not change except of the abovementioned tumors. Serum and tissue zinc level reduction was evident to certain extent in majority of tumors. Although insignificant differences were found, the analysis indicates that none of the tumors clearly disproves that the zinc levels remained unchanged. Variation of serum zinc level were found in esophageal cancer patients, in cell zinc content in liver cancer and both in serum and tissue zinc level in stomach, colorectal, and thyroid cancers.

Number of studies point to decreasing trend in tumors of higher grades or stages. Nevertheless, meta-regression could not be performed on the majority of tumors due to limited number of studies reporting stage/grade or to inconsistency in the scale used. Regression analysis of all tumors, however, did not show dependence on these parameters. Thus, this meta-analysis fails to explain the sources of high heterogeneity between the studies.

Although serum zinc level decrease in lung, head and neck, and breast carcinomas was shown by meta-analysis, it is unclear, whether hypozincaemia is a consequence of tumor, chronic stress or of a combination of both these effects. Stress, infection or chronic diseases lead to redistribution of zinc(II) between body compartments, and thus reduce zincaemia [126]. In addition, chronic inflammation is a common hallmark of cancer, and thus might be important mechanism of serum zinc level decrease.

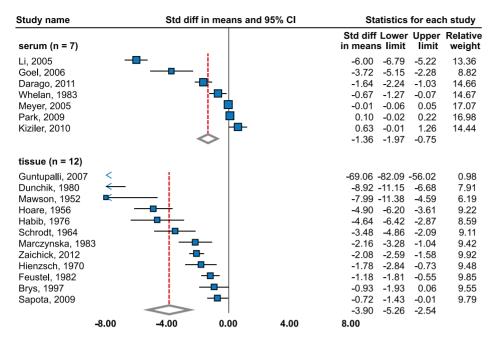


Figure 5. Zinc level in sera and tissues of lung cancer patients. Random effects model meta-analysis. Studies sorted by standardized mean difference.

doi:10.1371/journal.pone.0099790.g005

The association of tissue zinc level and prostate [6], liver [7], and lung [8] cancers serum zinc level and risk of breast, lung, stomach, and prostate cancers [9] were in scope of several reviews. Decrease in prostate cancer tissue zinc level is well-evidenced [127]. Also review by Zaichick et al. show decrease of zinc in prostate cancer tissue as compared with benign hyperplasia [6]. A review by Catalani, focusing on zinc content in lung tumors, is the only to date meta-analysis. However, its results did not allow summarizing the significance of tissue metals. No relationship among tissue zinc level and histotype or stage was found. Zinc decrease in liver cancer tissues were reviewed by Gurusamy et al. They declared that meta-analytic approach is impossible because of heterogeneity of analyzed studies. All mentioned reviews concluded that there is poor data agreement between studies determining tissue zinc level. This fact - combined with the low metal concentrations - calls for the standardization of methods. Catalani et al. propose standardization of sample collection, storage, and analysis. Previous reviews were performed only on specific tumors, with limited number of studies and/or statistic approaches were missing. Our meta-analytical analysis was done on all identified carcinomas, serum and tissue levels were analyzed together, publication bias was assessed and meta-regression was performed when case sufficient data were present. To reduce selection and publication biases, prospective cohort study with defined conditions separating the influence of inflammation is needed. Interest should be focused on the relation of zinc level in each histological type, stage, and grade.

There are limitations in this study caused by features of individual studies: sample sizes, subjects' characteristics, sampling, storage and detection methods, and different tumors classification.

References

- Maret W, Li Y (2009) Coordination Dynamics of Zinc in Proteins. Chem Rev 109: 4682–4707.
- Gumulec J, Masarik M, Krizkova S, Adam V, Hubalek J, et al. (2011) Insight to Physiology and Pathology of Zinc(II) Ions and Their Actions in Breast and Prostate Carcinoma. Curr Med Chem 18: 5041–5051.

Serum zinc level has a limited predictive value, because it is particularly intracellular ion and it fluctuates in circadian rhythm.

This meta-analysis shows a decrease of zincaemia in lung, head and neck, and breast carcinoma, increase of tissue zinc in breast cancer and its decrease in prostate, liver, and lung cancers. However, this analysis does not provide conclusive data with regard to stage and grade, and thus does not clarify heterogeneity in values between the studies.

Supporting Information

Checklist S1 Prisma 2009 checklist. (DOC)

Table S1 Meta-regression analysis of overall results using mixed effects model (unrestricted maximum likehood). SMD, standardized mean difference.

(XLSX)

Table S2 Source data set extracted from studies used for analysis. Including methodological quality of studies based on Newcastle-Ottawa scale, study design, and information regarding matching cases and controls. NS, not specified, NOS, Newcastle-Ottawa scale, stdev, standard deviation. (XLSX)

Author Contributions

Conceived and designed the experiments: JG VA MM RK. Performed the experiments: JG VA MM RK. Analyzed the data: JG IP TE. Contributed reagents/materials/analysis tools: RK MM IP. Wrote the paper: JG MM VA TE IP RK.

- Eckschlager T, Adam V, Hrabeta J, Figova K, Kizek R (2009) Metallothioneins and Cancer. Curr Protein Pept Sci 10: 360–375.
- Hogstrand C, Kille P, Nicholson RI, Taylor KM (2009) Zinc transporters and cancer: a potential role for ZIP7 as a hub for tyrosine kinase activation. Trends Mol Med 15: 101–111.

- Plum LM, Rink L, Haase H (2010) The Essential Toxin: Impact of Zinc on Human Health. Int J Environ Res Public Health 7: 1342–1365.
- Zaichick VY, Sviridova TV, Zaichick SV (1997) Zinc in human prostate gland: Normal, hyperplastic and cancerous. J Radioanal Nucl Chem 217: 157–161.
- Gurusamy K (2007) Trace element concentration in primary liver cancers A systematic review. Biol Trace Elem Res 118: 191–206.
- Catalani S, De Palma G, Mangili A, Apostoli P (2008) Metallic elements in lung tissues: results of a meta-analysis. Acta Biomed 79: 52–63.
- Silvera SAN, Rohan TE (2007) Trace elements and cancer risk: a review of the epidemiologic evidence. Cancer Causes Control 18: 7–27.
- Hess SY, Peerson JA, King JC, Brown KH (2007) Use of serum zinc concentration as an indicator of population zinc status. Food Nutr Bull 28: S403–S429.
- Wells GA, Shea B, O'Connell D, Peterson J, Welch V, et al. (2014) The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses.
- Higgins JPT, Thompson SG (2002) Quantifying heterogeneity in a metaanalysis. Stat Med 21: 1539–1558.
- Littell JH, Corcoran J, Pillai V (2008) Assessing Bias and Variations in Effects.
 In: Tripodi T, editor. Systematic Reviews and Meta-Analysis: Oxford University Press, USA. pp. 351.
- Higgins JPT, Green S (2011) Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0. In: Higgins JPT, Green S, editors: The Cochrane Collaboration.
- Boz A, Evliyaoglu O, Yildirim M, Erkan N, Karaca B (2005) The value of serum zinc, copper, ceruloplasmin levels in patients with gastrointestinal tract cancers. Turk J Gastroenterol 16: 81–84.
- Federico A, Iodice P, Federico P, Del Rio A, Mellone MC, et al. (2001) Effects
 of selenium and zinc supplementation on nutritional status in patients with
 cancer of digestive tract. Eur J Clin Nutr 55: 293–297.
- Chen H, Tan C, Wu T (2011) Ensemble Modeling Coupled with Six Element Concentrations in Human Blood for Cancer Diagnosis. Biol Trace Elem Res 143: 143–152.
- Kok FJ, Van Duijn CM, Hofman A, Van der Voet GB, De Wolff FA, et al. (1988) Serum copper and zinc and the risk of death from cancer and cardiovascular disease. Am J Epidemiol 128: 352–359.
- Leone N, Courbon D, Ducimetiere P, Zureik M (2006) Zinc, copper, and magnesium and risks for all-cause, cancer, and cardiovascular mortality. Epidemiology 17: 308–314.
- Zowczak M, Iskra M, Torlinski L, Cofta S (2001) Analysis of serum copper and zinc concentrations in cancer patients. Biol Trace Elem Res 82: 1–8.
- Gecit I, Kavak S, Demir H, Gunes M, Pirincci N, et al. (2011) Serum Trace Element Levels in Patients with Bladder Cancer. Asian Pac J Cancer Prev 12: 3409–3413.
- Mazdak H, Yazdekhasti F, Movahedian A, Mirkheshti N, Shafieian M (2010)
 The comparative study of serum iron, copper, and zinc levels between bladder cancer patients and a control group. Int Urol Nephrol 42: 89–93.
- Arinola OG, Charles-Davies MA (2008) Micronutrient levels in the plasma of Nigerian females with breast cancer. Afr J Biotechnol 7: 1620–1623.
- Cavallo F, Gerber M, Marubini E, Richardson S, Barbieri A, et al. (1991) Zinc and copper in breast cancer. A joint study in northern Italy and southern France. Cancer 67: 738–745.
- Garofalo JA, Ashikari H, Lesser ML, Menendezbotet C, Cunninghamrundles S, et al. (1980) Serum Zinc, Copper, and the Cu-Zn Ratio in Patients with Benign and Malignant Breast-Lesions. Cancer 46: 2682–2685.
- Gupta SK, Shukla VK, Vaidya MP, Roy SK, Gupta S (1991) Serum trace elements and Cu/Zn ratio in breast cancer patients. J Surg Oncol 46: 178–181.
- Huang Y-L, Sheu J-Y, Lin T-H (1999) Association between oxidative stress and changes of trace elements in patients with breast cancer. Clin Biochem 32: 131–136.
- Isbir T, Tamer L, Erkisi M, Kekec Y, Doran F, et al. (1995) Copper, Zinc and Magnesium in Serum and Tissues from Patients with Carcinoma of Breast, Stomach and Colon. Trace Elem Electrolytes 12: 113–115.
- Kuo HW, Chen SF, Wu CC, Chen DR, Lee JH (2002) Serum and tissue trace elements in patients with breast cancer in Taiwan. Biol Trace Elem Res 89: 1– 11.
- Magalova T, Bella V, Brtkova A, Beno I, Kudlackova M, et al. (1999) Copper, zinc and superoxide dismutase in precancerous, benign diseases and gastric, colorectal and breast cancer. Neoplasma 46: 100–104.
- Piccinini L, Borella P, Bargellini A, Medici CI, Zoboli A (1996) A case-control study on selenium, zinc, and copper in plasma and hair of subjects affected by breast and lung cancer. Biol Trace Elem Res 51: 23–30.
- Tinoco-Veras CM, Sousa MSB, da Silva BB, Cozzolino SMF, Pires LV, et al. (2011) Analysis of plasma and erythrocyte zinc levels in premenopausal women with breast cancer. Nutr Hosp 26: 293–297.
- Yucel I, Arpaci F, Ozet A, Doner B, Karayilanoglu T, et al. (1994) Serum copper and zinc levels and copper/zinc ratio in patients with breast cancer. Biol Trace Elem Res 40: 31–38.
- Carvalho ML, Magalhaes T, Becker M, von Bohlen A (2007) Trace elements in human cancerous and healthy tissues: A comparative study by EDXRF, TXRF, synchrotron radiation and PIXE. Spectroc Acta Pt B-Atom Spectr 62: 1004–1011.
- Ng KH, Bradley DA, Looi LM (1997) Elevated trace element concentrations in malignant breast tissues. Br J Radiol 70: 375–382.

- Raju GJN, Sarita P, Kumar MR, Murty G, Reddy BS, et al. (2006) Trace elemental correlation study in malignant and normal breast tissue by PIXE technique. Nucl Instrum Methods Phys Res Sect B-Beam Interact Mater Atoms 247: 361–367.
- Margalioth EJ, Schenker JG, Chevion M (1983) Copper and Zinc Levels in Normal and Malignant-Tissues. Cancer 52: 868–872.
- 38. Singh V, Garg AN (1998) Trace element correlations in the blood of Indian women with breast cancer. Biol Trace Elem Res 64: 237–245.
- Ebrahim AM, Eltayeb MAH, Shaat MK, Mohmed MA, Eltayeb EA, et al. (2007) Study of selected trace elements in cancerous and non-cancerous human breast tissues from Sudanese subjects using instrumental neutron activation analysis. Sci Total Environ 383: 52–58.
- Silva MP, Tomal A, Perez CA, Ribeiro-Silva A, Poletti ME (2009) Determination of Ca, Fe, Cu and Zn and their correlations in breast cancer and normal adjacent tissues. X-Ray Spectrom 38: 103–111.
- Geraki K, Farquharson MJ, Bradley DA (2002) Concentrations of Fe, Cu and Zn in breast tissue: a synchrotron XRF study. Phys Med Biol 47: 2327–2339.
- Farquharson MJ, Al-Ebraheem A, Geraki K, Leek R, Harris AL (2009) Zinc presence in invasive ductal carcinoma of the breast and its correlation with oestrogen receptor status. Phys Med Biol 54: 4213–4223.
- Stawarz R, Formicki G, Zakrzewski M, Rys J, Rozmus M (2009) Distribution of Heavy Metals and Trace Elements in Human Breast Cancer Tissues and in Adjacent Normal Tissues of Women in Poland. Fresen Environ Bull 18: 182– 188.
- Kubala-Kukus A, Banas D, Braziewicz J, Gozdz S, Majewska U, et al. (2007) Analysis of elemental concentration censored distributions in breast malignant and breast benign neoplasm tissues. Spectroc Acta Pt B-Atom Spectr 62: 695– 701.
- Ionescu JG, Novotny J, Stejskal V, Latsch A, Blaurock-Busch E, et al. (2006) Increased levels of transition metals in breast cancer tissue. Neuroendocrinol Lett 27: 36–39.
- Majewska U, Banas D, Braziewicz J, Gozdz S, Kubala-Kukus A, et al. (2007) Trace element concentration distributions in breast, lung and colon tissues. Phys Med Biol 52: 3895–3911.
- Cui Y, Vogt S, Olson N, Glass AG, Rohan TE (2007) Levels of zinc, selenium, calcium, and iron in benign breast tissue and risk of subsequent breast cancer. Cancer Epidemiol Biomarkers Prev 16: 1682–1685.
- Han CZ, Jing JX, Zhao ZW, Guo JG, Zheng SM, et al. (2003) Serum and tissue levels of six trace elements and copper/zinc ratio in patients with cervical cancer and uterine myoma. Biol Trace Elem Res 94: 113–122.
- Shobeiri MJ, Tabrizi AD, Atashkhoei S, Sayyah-Melli M, Ouladsahebmadarek E, et al. (2011) Serum levels of Copper, Zinc and Copper/Zinc Ratio in Patients with Ovarian Cancer. Pak J Med Sci 27: 561–565.
- Nasiadek M, Krawczyk T, Sapota A (2005) Tissue levels of cadmium and trace elements in patients with myoma and uterine cancer. Hum Exp Toxicol 24: 623-630
- Zhong HH, Tan MJ, Fu YL, Huang JQ, Tang ZF (1999) Determination of trace elements in tissue of human uterine cancer by instrumental neutron activation analysis. Biol Trace Elem Res 71–2: 569–574.
- Lin HJ, Chan WC, Fong YY, Newberne PM (1977) Zinc Levels in Serum, Hair and Tumors from Patients with Esophageal Cancer. Nutr Rep Int 15: 635–643.
- Goyal M, Kalwar A, Vyas R, Bhati A (2006) A study of serum zinc, selenium and copper levels in carcinoma of esophagus patients. Ind J Clin Biochem 21: 208–210.
- Mellow MH, Layne EA, Lipman TO, Kaushik M, Hostetler C, et al. (1983)
 Plasma zinc and vitamin a in human squamous carcinoma of the esophagus.
 Cancer 51: 1615–1620.
- Salehifar E, Khorasani G, Shokrzade M, Asadi M, Shabankhani B, et al. (2008)
 Comparison of plasma levels of zinc and lead in esophageal cancer patients with normal subjects. Trace Elem Electrolytes 25: 165–168.
- Abnet CC, Lai B, Qiao YL, Vogt S, Luo XM, et al. (2005) Zinc concentration in esophageal biopsy specimens measured by x-ray fluorescence and esophageal cancer risk. J Natl Cancer Inst 97: 301–306.
- Sun ZG, Song GM, Zhang M, Wang Z (2011) Clinical study on zinc, copper and manganese levels in patients with esophageal squamous cell cancer. Trace Elem Electrolytes 28: 116–120.
- 58. Kabuto M, Imai H, Yonezawa C, Neriishi K, Akiba S, et al. (1994) Prediagnostic serum selenium and zinc levels and subsequent risk of lung and stomach cancer in Japan. Cancer Epidemiol Biomarkers Prev 3: 465–469.
- Narang APS, Wani NA (1991) Levels of Copper and Zinc in Serum and Tissues of Patients with Gastric-Carcinoma. Trace Elem Med 8: 15–18.
- Yaman M, Kaya G, Yekeler H (2007) Distribution of trace metal concentrations in paired cancerous and non-cancerous human stomach tissues. World I Gastroenterol 13: 612–618.
- Milde D, Machacek J, Stuzka V (2007) Evaluation of colon cancer elements contents in serum using statistical methods. Chem Pap 61: 348–352.
- 62. Gupta SK, Shukla VK, Vaidya MP, Roy SK, Gupta S (1993) Serum and tissue trace elements in colorectal cancer. J Surg Oncol 52: 172–175.
- Zowczak-Drabarczyk M, Torlinska T, Iskra M, Mielcarz G, Matylla G, et al. (2004) Serum zinc concentration in patients with colorectal cancer. Trace Elem Electrolytes 21: 236–239.
- 64. Kim ES, Lim CS, Chun HJ, Keum B, Seo YS, et al. (2012) Detection of Cu(I) and Zn(II) ions in colon tissues by multi-photon microscopy: novel marker of antioxidant status of colon neoplasm. J Clin Pathol 65: 882–887.

- Hornik P, Milde D, Trenz Z, Vyslouzil K, Stuzka V (2006) Colon tissue concentrations of copper, iron, selenium, and zinc in colorectal carcinoma patients. Chem Pap-Chem Zvesti 60: 297–301.
- Kucharzewski M, Braziewicz J, Majewska U, Gozdz S (2003) Selenium, copper, and zinc concentrations in intestinal cancer tissue and in colon and rectum polyps. Biol Trace Elem Res 92: 1–10.
- Shukla VK, Adukia TK, Singh SP, Mishra CP, Mishra RN (2003) Micronutrients, antioxidants, and carcinoma of the gallbladder. J Surg Oncol 84: 31–35.
- Gupta SK, Singh SP, Shukla VK (2005) Copper, zinc, and Cu/Zn ratio in carcinoma of the gallbladder. J Surg Oncol 91: 204–208.
- Rautray TR, Vijayan V, Sudarshan M, Panigrahi S (2009) Analysis of blood and tissue in gallbladder cancer. Nucl Instrum Methods Phys Res Sect B-Beam Interact Mater Atoms 267: 2878–2883.
- Nagasue N, Kohno H, Chang YC, Nakamura T (1989) Iron, Copper and Zinc Levels in Serum and Cirrhotic Liver of Patients with and without Hepatocellular-Carcinoma. Oncology 46: 293–296.
- Poo JL, Rosas-Romero R, Montemayor AC, Isoard F, Uribe M (2003) Diagnostic value of the copper/zinc ratio in hepatocellular carcinoma: a case control study. J Gastroenterol 38: 45–51.
- Kolachi NF, Kazi TG, Afridi HI, Kazi NG, Khan S (2012) Investigation of essential trace and toxic elements in biological samples (blood, serum and scalp hair) of liver cirrhotic/cancer female patients before and after mineral supplementation. Clin Nutr 31: 967–973.
- Ebara M, Fukuda H, Hatano R, Yoshikawa M, Sugiura N, et al. (2003) Metal contents in the liver of patients with chronic liver disease caused by hepatitis C virus - Reference to hepatocellular carcinoma. Oncology 65: 323–330.
- Kew MC, Mallett RC (1974) Hepatic zinc concentrations in primary cancer of the liver. Br J Cancer 29: 80–83.
- Liaw KY, Lee PH, Wu FC, Tsai JS, LinShiau SY (1997) Zinc, copper, and superoxide dismutase in hepatocellular carcinoma. Am J Gastroenterol 92: 2260–2263.
- Maeda T, Shimada M, Harimoto N, Tsujta E, Maehara S, et al. (2005) Role of tissue trace elements in liver cancers and non-cancerous liver parenchyma. Hepato-Gastroenterol 52: 187–190.
- Okuno T, Shimamura Y, Mizuno M, Miyata S, Miyake T, et al. (1988) Trace-Elements in Hepatoma Tissue. Trace Elem Med 5: 130–136.
- Tashiro H, Kawamoto T, Okubo T, Koide O (2003) Variation in the distribution of trace elements in hepatoma. Biol Trace Elem Res 95: 49–63.
- Darago A, Sapota A, Matych J, Nasiadek M, Skezypinska-Gawrysiak M, et al. (2011) The correlation between zinc and insulin-like growth factor 1 (IGF-1), its binding protein (IGFBP-3) and prostate-specific antigen (PSA) in prostate cancer. Clin Chem Lab Med 49: 1699–1705.
- Goel T, Sankhwar SN (2006) Comparative study of zinc levels in benign and malignant lesions of the prostate. Scand J Urol Nephrol 40: 108–112.
- Kiziler AR, Aydemir B, Guzel S, Alici B, Ataus S, et al. (2010) May the level and ratio changes of trace elements be utilized in identification of disease progression and grade in prostatic cancer? Trace Elem Electrolytes 27: 65–72.
- Meyer F, Galan P, Douville P, Bairati I, Kegle P, et al. (2005) Antioxidant vitamin and mineral supplementation and prostate cancer prevention in the SU.VI.MAX trial. Int J Cancer 116: 182–186.
- Whelan P, Walker BE, Kelleher J (1983) Zinc, Vitamin A and Prostatic Cancer. Br J Urol 55: 525–528.
- 84. Li XM, Zhang L, Li J, Li Y, Wang HL, et al. (2005) Measurement of serum zinc improves prostate cancer detection efficiency in patients with PSA levels between 4 ng/mL and 10 ng/mL. Asian J Androl 7: 323–328.
- Park SY, Kolonel L (2009) Serum Zinc and Prostate Cancer Risk in the Multiethnic Cohort study. Epidemiology 20: S131–S131.
- Brys M, Nawrocka AD, Miekos E, Zydek C, Foksinski M, et al. (1997) Zinc and cadmium analysis in human prostate neoplasms. Biol Trace Elem Res 59: 145– 152.
- Dunchik VN, Zherbin EA, Zaichick VY, Leonov AI, Sviridova TV (1980) Method for differential diagnostics of malignant and benign prostate tumors. Discoveries, Inventions, Commercial Models, Trade Marks 35: 1–13.
- Feustel A, Wennrich R, Steiniger D, Klauss P (1982) Zinc and cadmium concentration in prostatic carcinoma of different histological grading in comparison to normal prostate tissue and adenofibromyomatosis (BPH). Urol Res 10: 301–303.
- Guntupalli JNR, Padala S, Gummuluri A, Muktineni RK, Byreddy SR, et al. (2007) Trace elemental analysis of normal, benign hypertrophic and cancerous tissues of the prostate gland using the particle-induced X-ray emission technique. Eur J Cancer Prev 16: 108–115.
- Habib FK, Hammond GL, Lee IR, Dawson JB, Mason MK, et al. (1976) Metal-androgen interrelationships in carcinoma and hyperplasia of the human prostate. J Endocrinol 71: 133–141.
- Marczynska A, Kulpa J, Lenko J (1983) The concentration of zinc in relation to fundamental elements in the diseased human prostate. Int Urol Nephrol 15: 257–265.
- 92. Mawson CA, Fischer MI (1952) The occurrence of zinc in the human prostate gland. Can J Med Sci 30: 336–339.
- Sapota A, Darago A, Taczalski J, Kilanowicz A (2009) Disturbed homeostasis
 of zinc and other essential elements in the prostate gland dependent on the
 character of pathological lesions. Biometals 22: 1041–1049.

- Schrodt GR, Hall T, Whitmore WF (1964) The concentration of zinc in diseased human prostate glands. Cancer 17: 1555–1566.
- Zaichick S, Zaichick V (2012) Trace elements of normal, benign hypertrophic and cancerous tissues of the human prostate gland investigated by neutron activation analysis. Appl Radiat Isot 70: 81–87.
- Hienzsch E, Schneider H, Anke M (1970) Vergleichende Untersuchungen zurn Mengenund Spurenelernentgehalt der normalen Prostata, des Prostataadenoms und des Prostatakarzinorns. Zsehr Urol 63: 543–546.
- 97. Hoare R, Delory GE, Penner DW (1956) Zinc and acid phosphatase in the human prostate. Cancer 9: 721–726.
- Garofalo JA, Erlandson E, Strong EW, Lesser M, Gerold F, et al. (1980) Serum zinc, serum copper, and the Cu/Zn ratio in patients with epidermoid cancers of the head and neck. J Surg Oncol 15: 381–386.
- Jayadeep A, Pillai KR, Kannan S, Nalinakumari KR, Mathew B, et al. (1997) Serum levels of copper, zinc, iron and ceruloplasmin in oral leukoplakia and squamous cell carcinoma. J Exp Clin Cancer Res 16: 295–300.
- Taysi S, Akcay F, Uslu C, Dogru Y, Gulcin I (2003) Trace elements and some extracellular antioxidant protein levels in serum of patients with laryngeal cancer. Biol Trace Elem Res 91: 11–18.
- 101. Akcil E, Caylakli F, Akiner M, Kocak M (2004) Trace element concentrations and superoxide dismutase and catalase activities in benign and malignant larynx tumors. Biol Trace Elem Res 101: 193–201.
- Drozdz M, Gierek T, Jendryczko A, Piekarska J, Pilch J, et al. (1989) Zinc, Vitamin-a and Vitamin-E, and Retinol-Binding Protein in Sera of Patients with Cancer of the Larynx. Neoplasma 36: 357–362.
- Durak I, Kavutcu M, Canbolat O, Isik AU, Akyol O (1994) Concentrations of Some Major and Minor Elements in Larynx Tissues with and without Cancer. Biometals 7: 45–48.
- 104. Yigitbasi OG, Haghighi N, Dogan P, Saraymen R, Balkani S, et al. (1998) Differences of tissue Cu and Zn levels in larynx squamous cell carcinomas. Trace Elem Electrolytes 15: 90–93.
- Al-Sayer H, Mathew TC, Asfar S, Khourshed M, Al-Bader A, et al. (2004) Serum changes in trace elements during thyroid cancers. Mol Cell Biochem 260: 1–5.
- 106. Przybylik-Mazurek E, Zagrodzki P, Kuzniarz-Rymarz S, Hubalewska-Dydejczyk A (2011) Thyroid Disorders-Assessments of Trace Elements, Clinical, and Laboratory Parameters. Biol Trace Elem Res 141: 65–75.
- Leung PL, Li XL (1996) Multielement analysis in serum of thyroid cancer patients before and after a surgical operation. Biol Trace Elem Res 51: 259– 266.
- Zaichick VY, Tsyb AF, Vtyurin BM (1995) Trace elements and thyroid cancer. Analyst 120: 817–821.
- 109. Kucharzewski M, Braziewicz J, Majewska U, Gozdz S (2003) Copper, zinc, and selenium in whole blood and thyroid tissue of people. with various thyroid diseases. Biol Trace Elem Res 93: 9–18.
- Homma S, Sasaki A, Nakai I, Sagai M, Koiso K, et al. (1993) Distribution of Copper, Selenium, and Zinc in Human Kidney Tumors by Nondestructive Synchrotron-Radiation X-Ray-Fluorescence Imaging. J Trace Elem Exp Med 6: 163–170.
- Dobrowolski Z, Drewniak T, Kwiatek W, Jakubik P (2002) Trace elements distribution in renal cell carcinoma depending on stage of disease. Eur Urol 42: 475–480.
- 112. Kwiatek WM, Drewniak T, Lekka M, Wajdowicz A (1996) Investigation of trace elements in cancer kidney tissues by SRIXE and PIXE. Nucl Instrum Methods Phys Res Sect B-Beam Interact Mater Atoms 109: 284–288.
- 113. Atukorala S, Basu TK, Dickerson JW, Donaldson D, Sakula A (1979) Vitamin A, zinc and lung cancer. Br J Cancer 40: 927–931.
- Klarod K, Hongsprabhas P, Khampitak T, Wirasorn K, Kiertiburanakul S, et al. (2011) Serum antioxidant levels and nutritional status in early and advanced stage lung cancer patients. Nutrition 27: 1156–1160.
- Diez M, Arroyo M, Cerdan FJ, Munoz M, Martin MA, et al. (1989) Serum and tissue trace metal levels in lung cancer. Oncology 46: 230–234.
- Oyama T, Matsuno K, Kawamoto T, Mitsudomi T, Shirakusa T, et al. (1994)
 Efficiency of serum copper/zinc ratio for differential diagnosis of patients with and without lung cancer. Biol Trace Elem Res 42: 115–127.
- 117. Beeley JM, Darke CS, Owen GCR (1974) Serum zinc, bronchiectasis, and bronchial carcinoma. Thorax 29: 21–25.
- Cildag O, Altinisik M, Kozaci D, Karadag F, Kiter G, et al. (2004) Alterations in trace elements and oxidative stress in lung cancer. Trace Elem Electrolytes 21: 23–27.
- Kune GA, Kune S, Watson LF, Pierce R, Field B, et al. (1989) Serum levels of beta-carotene, vitamin A, and zinc in male lung cancer cases and controls. Nutr Cancer 12: 169–176.
- 120. Sadat AFMN, Hossain MI, Hossain MK, Reza MS, Nahar Z, et al. (2008) Serum trace elements and immunoglobulin profile in lung cancer patients. J Appl Res 8: 24–33.
- 121. Jin Y, Zhang C, Xu H, Xue S, Wang Y, et al. (2010) Combined effects of serum trace metals and polymorphisms of CYP1A1 or GSTM1 on non-small cell lung cancer: A hospital based case-control study in China. Cancer Epidemiol 35: 182–187.
- 122. Sattar N, Scott HR, McMillan DC, Talwar D, O'Reilly DS, et al. (1997) Acute-phase reactants and plasma trace element concentrations in non-small cell lung cancer patients and controls. Nutr Cancer 28: 308–312.

- 123. Adachi S, Takemoto K, Ohshima S, Shimizu Y, Takahama M (1991) Metal concentrations in lung tissue of subjects suffering from lung cancer. Int Arch Occup Environ Health 63: 193-197.
- 124. Kubala-Kukus A, Braziewicz J, Banas D, Majewska U, Gozdz S, et al. (1999) Trace element load in cancer and normal lung tissue. Nucl Instrum Methods Phys Res Sect B-Beam Interact Mater Atoms 150: 193–199.
- 125. Zhang LL, Lv JG, Sun SK (2012) Elements in Lung Tissues of Patients from a
- High Lung Cancer Incidence Area of China. Biol Trace Elem Res 148: 7–10. 126. Cousins RJ, Liuzzi JP, Lichten LA (2006) Mammalian zinc transport, trafficking, and signals. J Biol Chem 281: 24085–24089.
- 127. Costello LC, Franklin RB (2011) Zinc is decreased in prostate cancer: an established relationship of prostate cancer! J Biol Inorg Chem 16: 3–8.