



Metallothionein Isoforms as Double Agents - Their Roles in Carcinogenesis, Cancer Progression and Chemoresistance

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1 **Metallothionein Isoforms as Double Agents - Their Roles in**
2 **Carcinogenesis, Cancer Progression and Chemoresistance**

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1 **Abstract**

2 Metallothioneins (MTs) are small cysteine-rich intracellular proteins with four major isoforms
3 identified in mammals, designated MT-1 through MT-4. The best-known biological functions
4 of MTs are their ability to bind and sequester metal ions and their active role in redox
5 homeostasis. Despite these protective roles, numerous studies have demonstrated that changes
6 in MT expression could be associated with the process of carcinogenesis and participation in
7 cell differentiation, proliferation, migration, and angiogenesis. Hence, MTs have the role of
8 double agents, *i.e.*, working with and against cancer. In view of their rich biochemical
9 properties, it is not surprising that MTs participate in the emergence of chemoresistance in
10 tumor cells. Many studies have demonstrated that MT overexpression is involved in the
11 acquisition of resistance to anticancer drugs, including cisplatin, anthracyclines, tyrosine
12 kinase inhibitors and mitomycin. The evidence is slowly becoming available for a cellular
13 switch in MT functions, showing that they indeed have two faces: protector and saboteur.
14 Initially, MTs display anti-oncogenic and protective roles; however, once the oncogenic
15 process has started, MTs are utilized by cancer cells for progression, survival, and the
16 coordination of chemoresistance. The duality of MTs can serve as a potential
17 prognostic/diagnostic biomarker and can therefore open the door to new strategies in cancer
18 treatment. Herein, we review and discuss MTs as tumor disease markers and describe their
19 role in chemoresistance to distinct anticancer drugs.

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24 **Keywords:** metallomics; tumor disease; drug resistance; cisplatin; anthracyclines, tyrosine
25 kinase inhibitors

1 **1. Introduction and main purpose of the review**

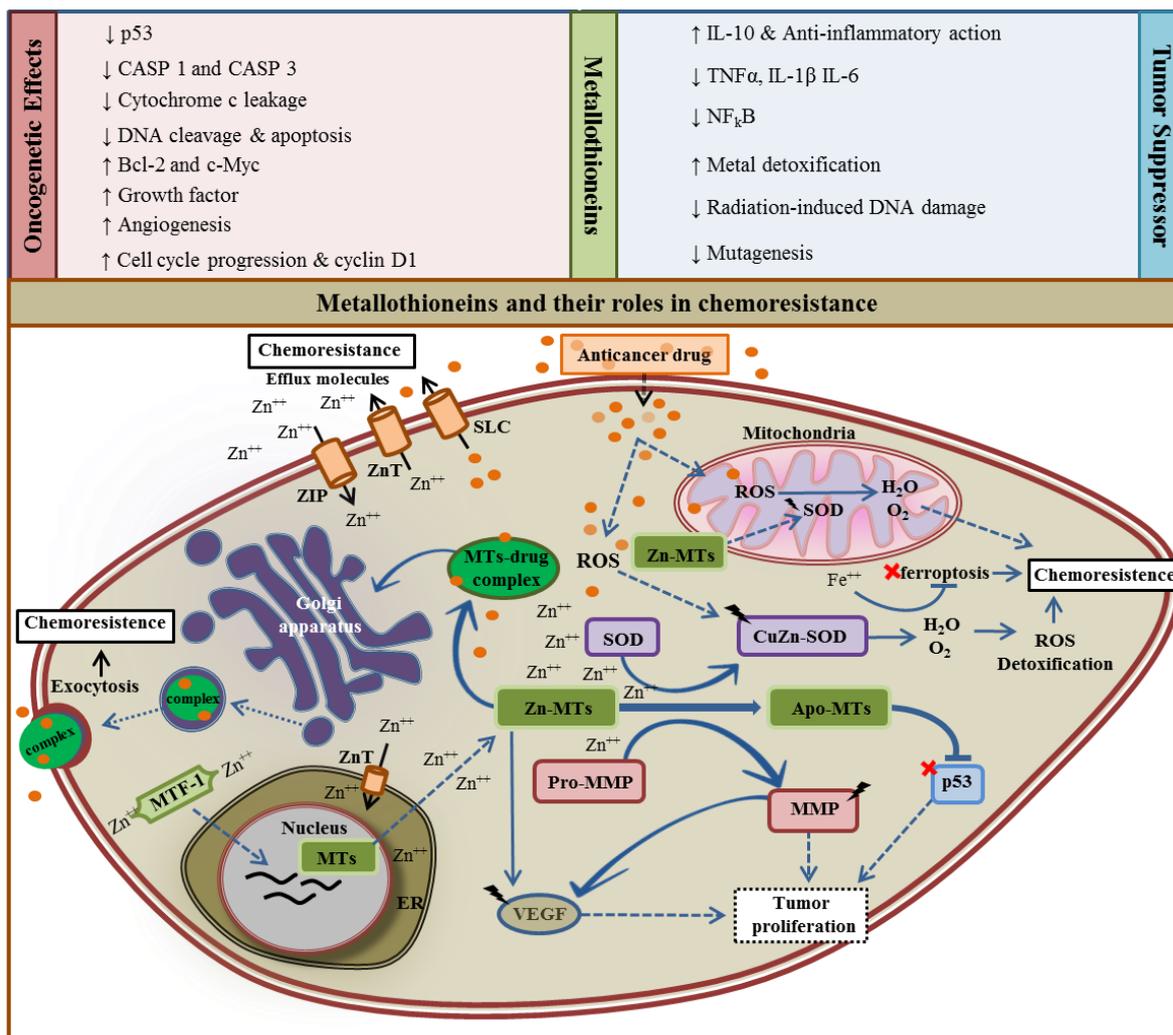
2 Metallothioneins (MTs) were discovered in 1957 by Margoshes and Vallee and identified as
3 low-molecular-mass sulfhydryl-rich proteins (Margoshes and Vallee, 1957). MTs have
4 molecular masses of 6 to 7 kDa and are characterized by an abundance of thiol groups (30%
5 cysteine) (Thirumoorthy et al., 2007). Interestingly, these major intracellular thiol-containing
6 proteins are induced by numerous agents, including UV radiation, DNA damaging agents, or
7 hormones and cytokines (Adam et al., 2016; Krizkova et al., 2012; Viarengo et al., 2000),
8 whose levels are elevated upon oxidative stress (Eckschlager et al., 2009; Ruttkay-Nedecky et
9 al., 2013).

10 In the last decade, it has been shown that MT overexpression is associated with
11 chemoresistance and poor prognosis in a variety of malignancies, particularly prostate, breast,
12 ovarian, head and neck cancer, non-small-cell lung cancer (NSCLC), melanoma,
13 neuroblastoma and soft tissue sarcoma (Chen et al., 2015; Hayden et al., 2014; Krizkova et
14 al., 2012; Lai et al., 2018; Lee et al., 2015; Raudenska et al., 2014; Si and Lang, 2018; Tariba
15 et al., 2015; Weinlich et al., 2003; Wong and Stillman, 2018). The currently accepted
16 mechanism of the role of MTs in chemoresistance is linked to their ability to chelate and
17 neutralize drugs, thus shielding vital biomolecules from the high reactivity and cytotoxic
18 effects of drugs and potentially leading to multidrug resistance (MDR) (Andrei et al., 2020;
19 Bar-Zeev et al., 2017; Coppola et al., 2017; Gacche and Assaraf, 2018; Gonen and Assaraf,
20 2012; Li et al., 2016a; Livney and Assaraf, 2013; Taylor et al., 2015; Wijdeven et al., 2016;
21 Zhitomirsky and Assaraf, 2016).

22 Therefore, it is not surprising that MT expression in tumor cells may be useful for
23 personalizing the treatment strategy. However, it is difficult to distinguish whether increased
24 MT expression is a factor inducing carcinogenesis and MDR or a factor inhibiting the
25 induction and development of cancer because of the irreplaceable protective roles of these

1 proteins in intracellular space. It should also be noted that increased MT expression has
2 protective cellular effects against carcinogenesis. On the other hand, the expression of MTs in
3 tumor cells protects them and increases the rate of tumor growth, resulting in decreased
4 patient survival (Masiulionyte et al., 2019; McGee et al., 2010). The anti-apoptotic functions
5 of MTs, (de)activation of transcription factors, and scavenging of reactive oxygen species
6 (ROS) are beneficial for cancer cell survival and proliferation and defense against the host
7 immune system, as depicted in **Figure 1** (Dutsch-Wicherek et al., 2008; Krizkova et al.,
8 2009b; Pedersen et al., 2009; Thirumoorthy et al., 2007). Thus, MTs can be considered
9 “double agents” that play crucial roles in both physiological processes and cancer.

10 In the last decade, numerous reviews have revealed and highlighted the importance of MTs as
11 a cancer biomarker; however, only a few of them were focused on the specific role of MTs in
12 drug chemoresistance (Bizon et al., 2017; Pedersen et al., 2009), since some of the most
13 important critical reviews were published more than two decades ago (Doz et al., 1993; Ebadi
14 and Iversen, 1994; Kelley et al., 1988). We therefore address this topic by highlighting not
15 only the current state of the art but also future directions paving new pathways towards
16 precision medicine strategies.



1

2 **Figure 1.** Involvement of MTs in various cancer-related processes: Oncogenesis (red), tumor
 3 suppressor (blue) and chemoresistance (brown) effects. MMP, matrix metalloproteinases; ER,
 4 endoplasmic reticulum; SOD, superoxide dismutase; p53, tumor protein p53; MTF-1, metal
 5 regulatory transcription factor; VEGF, vascular endothelial growth factor; ROS, reactive
 6 oxygen species; SLC, solute carrier group of membrane transport proteins; ZIP and ZnT, Zn⁺⁺
 7 transporter families.

8

9 **2. MT isoforms and structure**

10 Undoubtedly, it seemed incongruous to assume that a toxic metal would play a physiological
 11 role in a mammal when a cadmium-binding protein was first identified in horse kidney more
 12 than half a century ago (Margoshes and Vallee, 1957). The mystery of MTs has undergone
 13 many developments and continues to stir interest in many fields of pathophysiology including
 14 oxidative stress, metal toxicity, bone development, liver and kidney functions, heart disease,

1 diabetes, neurodegenerative disorders, cancer prognosis and, more recently, chemotherapy
2 development (Adam et al., 2016; Thirumoorthy et al., 2011).

3 Human MT isoforms are classified into four major groups: MT-1 and MT-2 are expressed in
4 many tissues, particularly in the liver and kidneys; MT-3 is mostly expressed in neurons and
5 male reproductive organs; and MT-4 is limited to the stratified squamous epithelium of the
6 skin and upper gastrointestinal tract (Klaassen et al., 1999; Thirumoorthy et al., 2011). Gene
7 expression profiling of MTs in human tissues revealed additional tissue-specific subgroups
8 among the MT-1 and MT-2 isoforms. In addition to the broadly expressed isoforms (MT-2A,
9 MT-1E, MT-1X, MT-1G, MT-1F and MT-1H), the following subtypes were clustered in
10 expression profiles with MT-3 and MT-4: MT-1A (highly expressed in intestine and adipose
11 tissue and moderately expressed in connective tissue, liver, lung eye and uterus), MT-1B
12 (highly expressed in connective tissue and moderately expressed in blood), and MT-1M
13 (moderately expressed in connective tissue, prostate, liver, lung intestine, uterus, stomach and
14 brain) (Moleirinho et al., 2011).

15 Metal-loaded MTs display a unique 3D structure consisting of two domains separated by a
16 short hinge region: the α -domain spans the C-terminus with a large number of cysteines (11
17 residues binding four Zn ions), forming a more rigid globule, while the β -domain spans the N-
18 terminal half, including a rather flexible cluster of 9 cysteine residues that bind three metal
19 ions (usually two Cd^{++} and one Zn^{++}) (Juarez-Rebollar et al., 2017; Klaassen et al., 1999).
20 There is, however, some uncertainty as to the role of the protonation of the thiol groups
21 themselves in MTs, as it is known that there are no disulfides. The protonation/deprotonation
22 of these cysteines is believed to be dynamic, with approximately 3 protonated thiols in each
23 domain that are constantly changing location (Szilagyi and Fenselau, 2000). Notably, MTs
24 also have several long-chained lysine residues that face the solution but may also extend
25 inward to neutralize free cysteines. The formation of intramolecular disulfides has been

1 reported, with higher frequency in the α -domain than the β -domain, and might play a
2 physiological role (Feng et al., 2006).

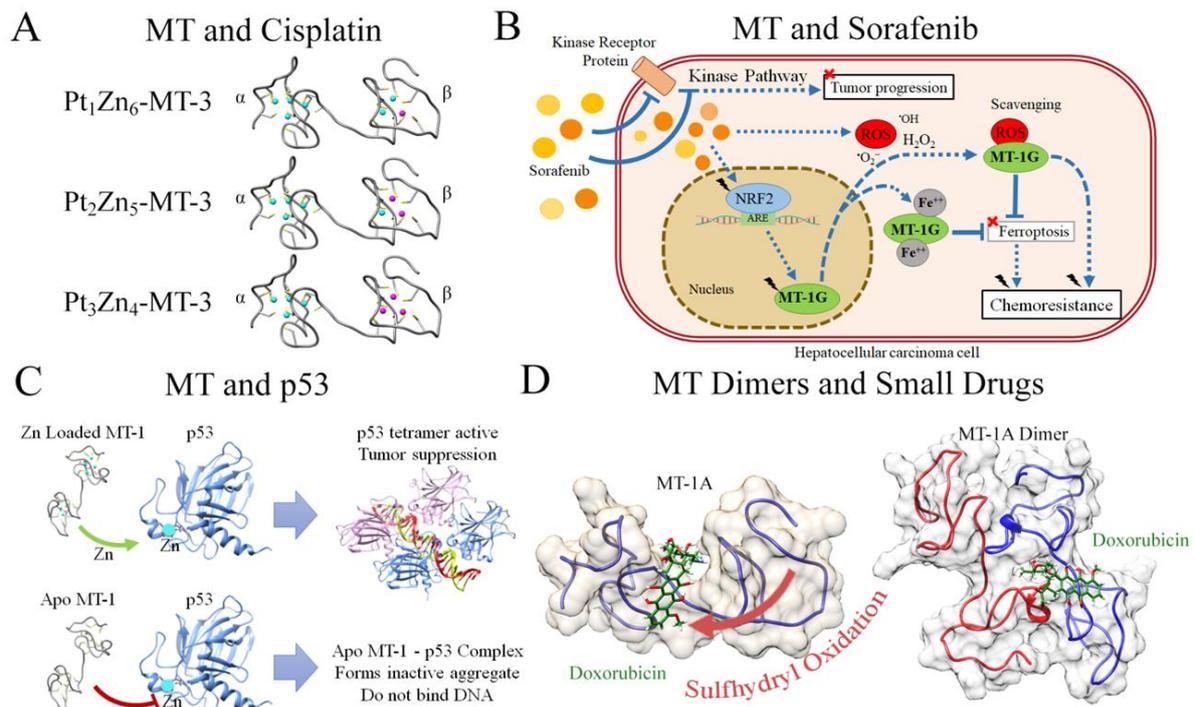
3 The above-described unique chemical properties determine the key roles of MTs in cellular
4 Zn^{++} homeostasis by low-affinity binding of approximately 5-15% of the cell's Zn ion pool in
5 combination with two classes of Zn transporter families, namely, Zrt- and Irt-like proteins
6 (ZIP, pumping Zn into cytoplasm) and Zn transporters (ZnT, pumping Zn away from
7 cytoplasm) (Golan et al., 2017; Kimura and Kambe, 2016). MT-3 was found to bind more
8 Cu^{++} than Zn^{++} ions in brain tissue compared to MT-1 and MT-2 (Adam et al., 2016; Artells
9 et al., 2014b). Another important and not yet well understood factor with a direct impact on
10 the function is MT dimerization and oligomerization. Further study is required to shed light
11 on these phenomena and their influence on MT binding affinity to metal ions and drugs
12 (Krizkova et al., 2009a; Ryvolova et al., 2012; Szilagy and Fenselau, 2000; Wilhelmsen et
13 al., 2002). MT oligomerization can occur either in native/nonoxidative forms (Artells et al.,
14 2014a) or in oxidative forms induced by high concentrations of Cd^{++} , where MT subunits are
15 covalently linked *via* disulfide bridges (Artells et al., 2014a; Zangger et al., 2001).

16 Interactions between MT isoforms and drugs can be either direct or indirect. An example of a
17 direct interaction is the neutralization of otherwise effective metal ion-based drugs such as
18 $Rh_2(AcO)_4$ by MT-1A (Wong and Stillman, 2016) and cisplatin by MT-3 (Kerotki and Vasak,
19 2009). In these cases, the metal ion replacement of Zn^{++} begins in the β -domain. MTs can
20 function as sensors and as transporters of metal ions through protein interactions. Cd^{++} -loaded
21 MT-1 and MT-2 isoforms bind directly to lipoprotein receptor-related proteins such as renal
22 megalin (LPR-2) *via* their hinge region SCKKSCC motif (Klassen et al., 2004). Indeed, many
23 interactions of metal-loaded MTs (and not apo-MTs) with specific proteins have been
24 reported (Atrian and Capdevila, 2013). Briefly, the binding of MTs (mainly MT-3) to the
25 transthyretin homotetramer enhances its ability to scavenge amyloid- β and prevent the onset

1 or progression of Alzheimer's disease (Adam et al., 2016). Similar claims have been reported
2 for α -synuclein protein in Parkinson's disease, for prions in spongiform encephalopathies, and
3 for many secreted transport proteins (*e.g.*, ferritin and albumin). In contrast, apo-MT-1
4 directly interacts with p53 to modulate Zn levels and p53 function. The p53 tumor suppressor
5 tetramer requires certain levels of Zn^{++} for proper folding of its DNA-binding domain (Lehvy
6 et al., 2019). This modulation has been proven by the inactivation of p53 in the presence of
7 highly expressed MT-1 and in the presence of apo-MT-1. On the other hand, sorafenib
8 upregulates MT-1G *via* an NRF2 transcription factor-dependent mechanism and not through
9 p53 or HIF-1 α (Sun et al., 2016). The upregulation of MT-1G contributes to sorafenib
10 chemoresistance in hepatocellular carcinoma (HCC) by inhibiting a form of nonapoptotic
11 regulated cell death called ferroptosis (involving glutathione depletion, lipid peroxidation and
12 iron metabolism). The MT structural interactions and mechanisms involved in
13 chemoresistance to various drugs are shown in **Fig. 2**.

14

15



1
 2 **Figure 2.** MT interactions and molecular mechanisms involved in drug chemoresistance. (A)
 3 The role of MT-3 in chemoresistance to cisplatin *via* replacement of Zn^{++} (cyan) with Pt
 4 (magenta). (B) Chemoresistance to sorafenib *via* activation of NRF2 transcription factor
 5 regulating the expression of MT-1G, which prevents ferroptosis by scavenging ROS resulting
 6 from iron metabolism (Fenton reaction). (C) Apo-MT-1 removes Zn^{++} (cyan) from p53 and
 7 forms an inactive complex, which prevents p53 from binding DNA and performing its
 8 function. (D) Inhibition of drugs by direct interaction with MTs. Reactive intermediates of
 9 doxorubicin (green) interact with mono- and dimer MT-1A *via* sulfhydryl oxidation. Crystal
 10 structures and homology models were used for these schematics (PDB ID: 3EXJ for p53 and
 11 4MT-2 for MT). UCSF Chimera (University of California San Francisco, CA, USA) was used
 12 for visualization and rendering.

13

14 3. MTs and cancer

15 Taking into account the abovementioned information, MTs are double agents because they
 16 can also control the homeostasis of Zn^{++}/Cu^{++} in cells, which is essential for proliferation and
 17 differentiation (Adam et al., 2016; Krizkova et al., 2018). The antioxidant function of MTs
 18 protects the cells from free radicals and oxidative stress arising from mutagens, anticancer
 19 drugs, and radiation. The ability of MTs to bind Cd^{++} , Hg^{++} , Pt and other similar heavy metals
 20 protects cells from the toxicity of these metals (Krizkova et al., 2012; McNeill et al., 2019;
 21 Rahman et al., 2017; Wong and Stillman, 2018). On the other hand, it has been demonstrated
 22 that changes in MT expression could be associated with the process of carcinogenesis and

1 cancer progression (Krizkova et al., 2018; Si and Lang, 2018). Herein, we summarize the
2 current findings associating MTs and cancer.

3 *3.1. MT involvement in carcinogenesis*

4 Carcinogenesis is a process of tumor formation by the transformation of normal cells into
5 cancer cells. This transformation originates from an abnormal “program” leading to a cascade
6 of downstream changes (Jones and Baylin, 2007). All these changes at the cellular, genetic,
7 and epigenetic levels disrupt the balance between proliferation and apoptosis (*e.g.*, mutations
8 and epimutations) and thus contribute to the development of cancer. MTs can promote tumor
9 growth by regulating the supply of Zn^{++} for proteins and the activity of Zn-dependent
10 transcription factors or by direct interaction with other proteins (Krizkova et al., 2018;
11 Krizkova et al., 2012; Zalewska et al., 2014). MTs are also involved in the cell cycle
12 regulation, cell proliferation, and the inhibition of apoptosis (Krizkova et al., 2009b; Si and
13 Lang, 2018). It was previously observed that the cytoplasmic levels of MTs reached a
14 maximum during the G_1/S phase of the cell cycle (Nagel and Vallee, 1995) and that Zn^{++} is
15 required for the G_1/S phase transition (He et al., 2019). Werynska et al. demonstrated the
16 significance of MT-1/2 expression in the pathogenesis of lung adenocarcinoma. MT-1/2
17 expression was shown in proliferating NSCLC cells, pointing to the prognostic importance of
18 the parallel expression of MT-1/2 and Ki-67, which are manifested mainly in the late G_1 , S,
19 G_2 and M phases of the cell cycle. Ki-67 is one of the most frequently employed markers of
20 cell proliferation (Werynska et al., 2011). Similar correlations were found in breast cancer
21 (Gomulkiewicz et al., 2010; Jin et al., 2002), nasopharyngeal carcinoma (Jayasurya et al.,
22 2000), colon adenocarcinoma (Dziegiel et al., 2003), basal cell carcinoma (Bieniek et al.,
23 2012), and soft tissue sarcomas, such as malignant fibrous histiocytoma, liposarcoma, and
24 synovial sarcoma (Dziegiel et al., 2005). MTs can transfer Zn^{++} to transcription factors such
25 as HIF-1 α and tumor suppressors such as p53 (Krizkova et al., 2012) and have been found to

1 interact with NF- κ B to mediate its anti-apoptotic effect (Krizkova et al., 2009b). MT
2 overexpression is consistently associated with the presence of mutant p53 in breast cancer,
3 anti-apoptotic effects, differentiation, proliferation, progression and poor prognosis (Sampaio
4 et al., 2019). The interaction between MT-2A and Fas-associated protein with the death
5 domain was connected with increased cell proliferation and apoptosis inhibition in colorectal
6 cancer (CRC) *via* the NF- κ B pathway (Marikar et al., 2016). Furthermore, MT expression can
7 also protect cancer cells against a variety of pro-apoptotic stimuli, such as chemotherapeutics,
8 heavy metals, oxidative stress, and radiation. For a detailed description, see the following
9 sections.

10 Thus far, we do not know the underlying molecular mechanisms that explain why MT
11 expression is increased in some cancers and downregulated in others. The alterations in the
12 expression patterns of isoforms could be a possible explanation. Accordingly, the
13 upregulation of specific MT isoforms was found to affect the growth of low-MT-expressing
14 cancer cells. The transfection of MT-1F into CRC cells decreased their proliferation, colony
15 formation and increased apoptosis (Yan et al., 2012). Low expression and tumor suppressor
16 activity of MT-1H were found in prostate cancer cells (Zheng et al., 2017). The expression of
17 MT-3 in PC-3 cells reduced cell growth (Dutta et al., 2002).

18 *3.2. MTs in tumor differentiation and angiogenesis*

19 Cellular differentiation is essential for tissue and organ development. Undifferentiated or
20 poorly differentiated cells are more likely than differentiated cells to form tumors. Cancer is
21 characterized by the grade of histological differentiation, which is used to determine cancer
22 progression. Multiple studies have reported the participation of MTs in cell differentiation;
23 however, these roles are also isoform-specific. The influence of extremely low-frequency
24 electromagnetic fields on Zn⁺⁺-MT-3 interaction during neuronal differentiation was studied.
25 During this interaction, the expression of MT-3 was downregulated and the formation of

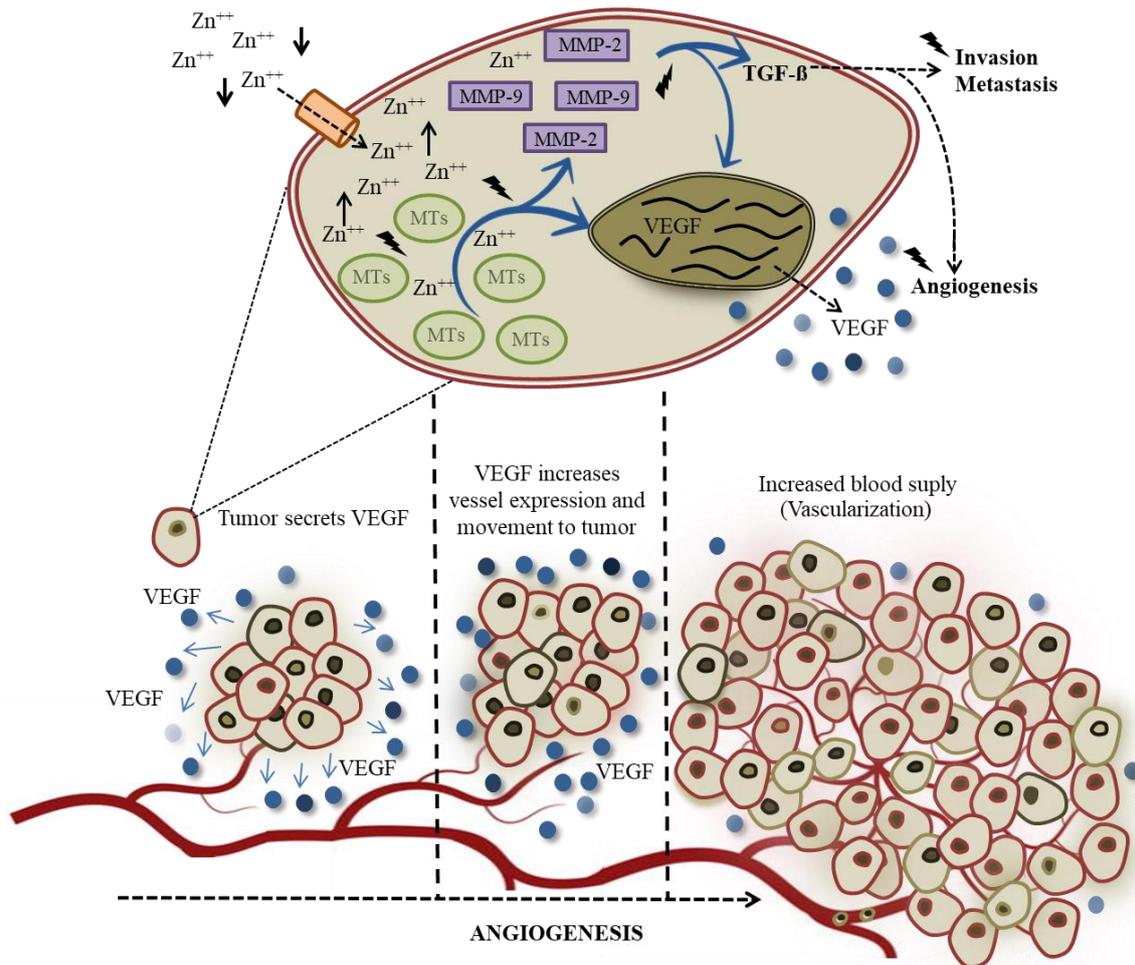
1 Zn^{++} -MT-3 complexes was increased to maintain Zn^{++} homeostasis (Aikins et al., 2017). MTs
2 were also found to negatively regulate IL-27-induced type 1 regulatory T-cell differentiation
3 (Wu et al., 2013). MT-1G overexpression inhibited the all-trans retinoic acid-induced
4 differentiation of NB-4 cells (Hirako et al., 2014). MT-2A overexpression enhanced the
5 differentiation of osteosarcoma cells towards the osteogenic lineage (Habel et al., 2013). The
6 expression of M-1F and MT-2A in histological grade 3 breast cancer was significantly
7 increased compared to grades 1 and 2 (Jin et al., 2001; Jin et al., 2002). Similar results have
8 also been published for ductal breast cancers, indicating connections to the chemoresistance,
9 invasiveness, and clinical stage of breast cancers (Gallicchio et al., 2005; Gomulkiewicz et al.,
10 2010; Rezk et al., 2015; Yap et al., 2009). The relationship between MT expression and tumor
11 histological grade was also demonstrated in pancreatic ductal carcinoma (Ohshio et al., 1996),
12 gallbladder carcinoma (Shukla et al., 1998), renal cancer (Mitropoulos et al., 2005), ovarian
13 adenocarcinoma (Hengstler et al., 2001; McCluggage et al., 2002), and endometrial
14 carcinoma (Bredholt et al., 2015). MT-1G was also found to be involved in the differentiation
15 of CRC cells through the Notch signaling pathway and labile Zn^{++} chelation and redistribution
16 (Arriaga et al., 2017).

17 MTs can induce the upregulation of angiogenesis-related genes, such as matrix
18 metalloproteinases (*MMP-9* and *MMP-2*) and *VEGF*, to form new blood vessels (**Figure 3**).
19 This phenomenon is an important step in tumorigenesis to supply the tumor with oxygen and
20 nutrients for its growth, progression and metastasis. MMPs are associated with tumor
21 progression because of their role in remodeling of the extracellular matrix, angiogenesis and
22 revascularization (Cho et al., 2019). For instance, *MMP-9* (*i.e.*, gelatinase B) was found to
23 interact directly with MTs (Zalewska et al., 2014). Following MT knockout, dysfunction of
24 endothelial cells (ECs), smooth muscle cells and macrophages was observed in mice. *MMP-9*,

1 PDGF, and VEGF were significantly downregulated in these mice, which contributed to these
2 dysfunctions (Zbinden et al., 2010).

3 MT-1 was found to be expressed in vascular endothelial cells at the site of angiogenesis, and
4 its downregulation resulted in inhibited cell proliferation, migration and angiogenesis *in vivo*
5 (Miyashita and Sato, 2005). Decreased levels of the growth factors β -FGF, TGF- β 1, and
6 VEGF mediated decreased angiogenesis and regeneration in MT-1/2-deficient mice after
7 cortical freeze injury. These mice also displayed a dramatic reduction in IL-6-induced
8 angiogenesis (Miyashita and Sato, 2005; Penkowa et al., 2000). The expression of VEGF was
9 slightly increased in breast cancer cell lines after exposure to Zn⁺⁺ ions, which also led to
10 increased expression of selected MT isoforms. These results suggested a correlation between
11 MTs and VEGF expression in these cell lines (Wierzowiecka et al., 2016). In brain ECs, MT-
12 3 was found to induce the expression of VEGF through an HIF-1 α -dependent mechanism
13 (Kim et al., 2008). MT-2 was also found to induce the expression of VEGF in the regulation
14 of EC proliferation, migration and angiogenesis (Schuermann et al., 2015).

15



1

2 **Figure 3.** Roles of MTs in tumor angiogenesis. Zn⁺⁺ compartmentalization within the tumor
 3 favors invasion, metastasis, and angiogenesis mediated by MTs and matrix metalloproteinases
 4 (MMP-2 and MMP-9). MTs and MMPs act upstream of VEGF and TGF-β in regulating
 5 angiogenesis. In turn, these new blood vessels supply the growing tumors with oxygen and
 6 nutrients, allowing the cancer cells to invade nearby tissue, thereby spreading throughout the
 7 bloodstream and culminating in metastasis.

8

9 *3.3. The role of MTs in tumor metastasis*

10 During cancer development, the stage of tumor metastasis is reached when cells from the
 11 primary tumor undergo dissemination to a secondary site. Some studies have shown that MTs
 12 collaborate with invasion, spread, and metastasis in various types of cancers, so they can be
 13 used as markers in aggressive cancers (Pedersen et al., 2009; Si and Lang, 2018). MTs may
 14 have a significant role in oncogenesis, but their expression is divergent in different kinds of

1 human tumors, and it is related to tumorigenesis and tumor progression (Si and Lang, 2018).
2 Moreover, MT isoforms were also identified in numerous benign lesions with different
3 expression patterns in these tissues (Fic et al., 2013; Krizkova et al., 2012; Pula et al., 2015).
4 As mentioned earlier, a few studies reported that MT expression was downregulated in some
5 cancers (*e.g.*, gastric, HCC and CRC), while in other cancers, it was upregulated (breast,
6 kidney, lung and prostate). Hence, MT expression is associated with the type and status of the
7 tumors and might be a useful tool for the choice of therapy or for cancer diagnosis (Kim et al.,
8 2011). For example, MT overexpression (MT-1/2) was correlated with tumor-infiltrating
9 macrophages (a known predictive value of progression and correlated with metastasis
10 formation) in cutaneous malignant melanoma, as previously reported by Emri et al. (Emri et
11 al., 2013). Furthermore, Suzuki et al. suggested that these two MTs have protective values in
12 the initial stages of skin carcinogenesis (Suzuki et al., 2003), accelerating wound healing in
13 keratinocytes (Morellini et al., 2010), but once carcinogenesis occurs, MTs promote tumor
14 growth (McGee et al., 2010).

15 MT-1 and MT-2 operate as protective agents against *Helicobacter pylori* infection (which
16 over time may result in the development of gastric cancer) (Tran et al., 2007). However, MT-
17 2 has also been identified as a potent tumor suppressor that prevents the development of
18 gastric cancer (Pan et al., 2013a; Pan et al., 2016; Pan et al., 2013b). Decreased expression of
19 MT-2 and MT-3 was shown in gastric cancer specimens compared to healthy gastric mucosa
20 (Deng et al., 2003; Ebert et al., 2000). Additionally, MT overexpression was correlated with
21 lymph node metastasis in gastric cancers (Galizia et al., 2006). Other studies showed low
22 expression of MT-1E, MT-1F, MT-1G, MT-1H, MT-1M, MT-1X, and MT-2A isoforms in
23 the CRC mucosa compared with normal mucosa, suggesting that these MT isoforms are
24 potent tumor suppressors (Fic et al., 2013; Janssen et al., 2000; Pedersen et al., 2009; Yan et
25 al., 2012). In HCC samples, some molecular studies confirm the specific downregulation of

1 MT isoforms, including *MT-1A*, *MT-1E*, *MT-1G*, *MT-1M*, *MT-1H*, *MT-1X*, and *MT-2*, with no
2 differences in regard to *MT-1B*, *MT-1E*, or *MT-1X* (Datta et al., 2007; Mao et al., 2012). In
3 addition, the analysis of patients' clinical and pathological data revealed that MT expression
4 was inversely correlated with the malignancy grade of the tumors and the clinical
5 advancement stage of cancer (Endo et al., 2003; Endo et al., 2004; Tao et al., 2007). *MT-1G*
6 downregulation has also been observed in hepatoblastoma (Sakamoto et al., 2010).
7 Furthermore, the transcript levels of *MT-2* were significantly upregulated in renal cancer,
8 whereas *MT-1A* and *MT-1G* were downregulated (Nguyen et al., 2000). The *MT-3* gene was
9 among the reported downregulated metastasis genes involved in primary solid tumors in this
10 type of cancer (Fu et al., 2013).

11 In a study of HCC cells, Lui et al. demonstrated that *MT-1X* overexpression delayed the cell
12 cycle and promoted apoptosis *in vitro* and *in vivo*. Moreover, *MT-1X* suppressed tumor
13 growth and promoted lung metastasis. The study also demonstrated that *MT-1X* caused the
14 inactivation of NF- κ B signaling, thus resulting in cell cycle arrest and apoptosis (Liu et al.,
15 2018).

16 MT expression in normal lung tissue seems to protect against dangerous factors that can
17 induce oncogenesis (Inoue et al., 2008; Takaishi et al., 2010). However, lung epithelial cells
18 with a high level of MTs may contribute to the later induction of oncogenesis (Person et al.,
19 2013). Thus, it seems that MTs protect lung cells from damaging factors until some critical
20 event; however, once carcinogenesis begins, they contribute to tumor progression (McGee et
21 al., 2010; Werynska et al., 2011).

22 MTs are also involved in the oncogenesis of head and neck carcinomas. Hishikawa et al.
23 showed that MT expression in squamous cell carcinoma of the head and neck was positively
24 correlated with metastasis and tumor cell proliferation in the esophagus (Hishikawa et al.,
25 1997). Brazao-Silva et al. demonstrated a different behavior of MTs in normal oral mucosa

1 and oral squamous cell carcinoma. Both tissues displayed the same expression of *MT-1B* and
2 *MT-1H*. On the other hand, oral squamous cell carcinoma exhibited downregulation of *MT-*
3 *1A*, *MT-1X*, *MT-3* and *MT-4*, in contrast to upregulation of *MT-1F* expression. Moreover, the
4 patients with the worst prognosis exhibited downregulation of *MT-1G*, while *MT-1X*
5 overexpression was observed in nonmetastatic cases, in contrast to *MT-3* overexpression,
6 which was observed solely in the patients who presented with metastasis (Brazao-Silva et al.,
7 2015). Increased expression of MT-1 and MT-2 in cervical cancer cells has been
8 demonstrated previously. MT-1/2 expression is prominent in the basal layer cells of the
9 normal uterus, in cervical intraepithelial neoplasia, and during tumor progression to stage II–
10 III (Theocharis et al., 2004). Endometrial carcinomas also display an increase in MT-1/2 that
11 is positively correlated with higher tumor grade, higher tumor cell proliferative capacity, and
12 reduced patient survival (Dutsch-Wicherek et al., 2008; Theocharis et al., 2004). Furthermore,
13 Fu et al. demonstrated a significant positive association of *MT-1G* hypermethylation with
14 lymph node metastasis in 178 papillary thyroid cancer patients (Fu et al., 2013).

15 Several studies have also concluded that the expression of distinct MT isoforms was
16 associated with poor survival, tumor grade, and recurrence rate in highly malignant invasive
17 ductal breast carcinomas (Bay et al., 2006; Cherian et al., 2003; Dziegiel et al., 2004;
18 Gallicchio et al., 2005; Surowiak et al., 2006). All isoforms of MT-1, MT-2 and MT-3 have
19 been detected in breast cancer tissue samples (Jin et al., 2002; Jin et al., 2004; Sens et al.,
20 2002). Moreover, different expression patterns of *MT-1* have been noted in cases of breast
21 cancer with confirmed lymph node metastases (Dutsch-Wicherek et al., 2005). MT-1F has
22 been positively correlated with malignancy grade (Jin et al., 2001), whereas *MT-1E*
23 expression has been found in estrogen receptor-negative breast cancer cell lines (Friedline et
24 al., 1998; Jin et al., 2001). In addition, *MT-2A* has been identified as the most abundant in
25 breast cancer and plays a protective role in these cancer cells, with high expression involved

1 in cell cycle regulatory capabilities (Lai et al., 2010; Lai et al., 2011). MT overexpression can
2 also induce other proteins, *e.g.*, MT-2A induces the overexpression of MMP-9 with
3 subsequent induction of the invasive phenotype of breast cancer cells (Emri et al., 2013). In
4 numerous cases of breast cancer, the expression status of estrogen and progesterone receptors
5 was inversely correlated with MT-1 and MT-2 expression (El Sharkarvy and Farrag, 2008;
6 Gomulkiewicz et al., 2010).

7 In normal prostate tissue, differential MT expression was observed, namely, high expression
8 in low-grade cancers and lack of expression in high-grade cancers (Wei et al., 2008).
9 Gumulec et al. found differential expression of MTs in prostate cancer at the RNA and protein
10 levels, particularly an increase in MT-1A-encoding mRNA levels and a simultaneous
11 decrease in the MT-1/2 proteins (Gumulec et al., 2012). In addition, Han et al. demonstrated
12 that low expression of *MT-1H* attenuated cell migration and cell growth (Han et al., 2013). On
13 the other hand, *MT-1G* hypermethylation has been linked to the aggressiveness of lesions and
14 is characteristic of high-grade prostate cancers (Henrique et al., 2005). In summary, MTs are
15 involved in tumor growth, differentiation, and metastasis. The up- and downregulation of
16 MTs contributes to metastasis and increases the adhesion, invasion, and migration of tumor
17 cells. This variation in expression behavior depends on the MT isoforms and the tumor type,
18 as well as on the tumor microenvironment (Theocharis et al., 2004).

19 3.4. *MTs as cancer biomarkers*

20 MTs manifest varying expression levels in carcinomas and thus may serve as valuable cancer
21 biomarkers in certain malignances. An interesting meta-analysis study was conducted to
22 determine the characteristics of MT expression in benign and malignant tumors originating
23 from different tissues. The results of immunohistochemical evaluation of MTs in benign
24 tumors revealed an important downregulation, in contrast to malignant tumors, particularly
25 since this difference appeared in many tumors (Zhang et al., 2014). In addition, Gumulec et

1 al. evaluated the associations between MT expression and clinicopathological characteristics,
2 tumor type, stage, grade, prognosis, and survival using a meta-analysis. While no associations
3 were identified between MTs and tumor staging, a positive association was found with tumor
4 grade. In particular, strong associations were observed in breast, ovarian, uterine and prostate
5 cancers (Gumulec et al., 2014b). There are currently a number of good systematic reviews of
6 MTs as biomarkers for cancer diagnosis (Babula et al., 2012; Bizon et al., 2017; Cherian et
7 al., 2003; Dziegiel et al., 2016; Felizola et al., 2014; Gumulec et al., 2012; Jin et al., 2004;
8 Krizkova et al., 2009b; Krizkova et al., 2012; Malavolta et al., 2016; Miles et al., 2000; Si and
9 Lang, 2018; Takahashi, 2012). Associations of MTs with several diseases, such as
10 Alzheimer's disease (Adam et al., 2016; Roy et al., 2017; Waller et al., 2018), circulatory
11 diseases (Billaud et al., 2018; Cong et al., 2016; Yu et al., 2018) and amyotrophic lateral
12 sclerosis (Ono, 2017), have also been found. Furthermore, strong evidence exists regarding
13 the potential role of MTs in the immune system and inflammatory processes (Pankhurst et al.,
14 2011; Waeytens et al., 2009; Youn et al., 2002).

15 **4. MTs in chemoresistance to anticancer drugs**

16 Chemoresistance, a complex system of heterogeneous biochemical mechanisms, is mainly
17 embodied in the insensitivity of cancer cells to therapy and is considered a key factor in the
18 failure of anticancer chemotherapy (Andrei et al., 2020; Bar-Zeev et al., 2017; Coppola et al.,
19 2017; Gacche and Assaraf, 2018; Gonen and Assaraf, 2012; Hanahan and Weinberg, 2011;
20 Kopecka et al., 2020; Li et al., 2016a; Livney and Assaraf, 2013; Taylor et al., 2015;
21 Wijdeven et al., 2016; Zhitomirsky and Assaraf, 2016). In view of the rich biochemical
22 properties of MTs, it is not surprising that they are believed to participate in the emergence of
23 chemo- and/or radioresistance in tumor cells. It has been suggested that MTs provide
24 protection against apoptosis and promote cell proliferation, leading to tumorigenesis
25 (Krzeslak et al., 2014). Drug resistance has been postulated to be mainly the result of

1 protection against ROS and anti-apoptotic factors and of the direct sequestration of alkylating
2 agents by MT cysteines (Habel et al., 2013; Lai et al., 2011). Recent evidence also supports
3 interactions with other important thiol compounds, indicating the involvement of glutathiones
4 in chemoresistance (Kim et al., 2019; Russi et al., 2019; Tanner et al., 2002); however, a
5 direct mechanism of interaction has not yet been elucidated. **Table 1** shows an overview of
6 the upregulated expression of MTs in chemoresistance to anticancer drugs. As shown by Yap
7 et al., siRNA-based silencing of *MT-2A* in MCF-7 cells exposed to doxorubicin (Dox) led to a
8 significant reduction in cell viability and a corresponding increase in apoptosis (Yap et al.,
9 2009). Similarly, poor survival was observed in bladder tumors expressing higher levels of
10 MTs due to their mediation of resistance against alkylating agents (Wulfing et al., 2007). MTs
11 were also shown to initiate Dox resistance in NSCLC (Mattern and Volm, 1992), in which a
12 significant relationship between MT expression and resistance was found. Moreover, a
13 significant correlation was also documented between MT and glutathione *S*-transferase P
14 enzyme expression. However, the role of MTs in the development of chemoresistance in
15 clinical conditions is still controversial, and their importance may vary in different tumors.

16 **Table 1.** Overview of the upregulated expression of MTs in chemoresistance to anticancer
17 drugs.

| Cancer type | MT isoforms | Chemotherapy | References |
|----------------------|---|----------------------|--|
| Testicular cancer | Total MTs | cisplatin | (Chin et al., 1993; Tariba et al., 2015) |
| Esophageal cancer | Total MTs | cisplatin | (Hishikawa et al., 1997; Hou et al., 2017) |
| Urothelial carcinoma | MT-1A, MT-1B | cisplatin | (Skowron et al., 2018) |
| Neuroblastoma | MT-3 | cisplatin | (Rodrigo et al., 2018) |
| Bladder cancer | MT-2A | mitomycin, cisplatin | (Lynn et al., 2003; Singh et al., 1995) |
| | Total MTs | cisplatin | (Hayden et al., 2014; Kondo et al., 1992; Wulfing et al., 2007) |
| Prostate cancer | MT-2A, MT-1E, MT-1G, MT-1R, MT-1 L, MT-3, Total MTs | cisplatin | (Dutta et al., 2002; Gumulec et al., 2014a; Henrique et al., 2005; Smith et al., 2006) |
| | MT-1E, MT3 | mitoxantrone | (Dutta et al., 2002) |
| | MT-1, MT-2, MT1-E, MT-3 | cadmium, arsenic | (Dutta et al., 2002; Lee et al., 1999) |

| | | | |
|--------------------------------|-----------------------------------|-----------------|--|
| | MT-1E, MT3 | etoposide | (Dutta et al., 2002) |
| | MT-1E, MT3 | vinblastine | (Dutta et al., 2002) |
| | MT-1E, MT3 | paclitaxel | (Dutta et al., 2002) |
| | MT1-X | genistein | (Raschke et al., 2006) |
| Gastric cancer | MT-1G | docetaxel | (Pan et al., 2016) |
| | MT-1X | irinotecan | (Chun et al., 2004a) |
| | MT-1G | cisplatin | (Suganuma et al., 2003) |
| NSCLC | MT-1H | doxorubicin | (Mattern and Volm, 1992) |
| Ovarian cancer | Total MTs, MT-2A | cisplatin | (Andrews et al., 1987; Cheng et al., 2006; Surowiak et al., 2005) |
| CRC | MT-1G | oxaliplatin | (Arriaga et al., 2017; Arriaga et al., 2014) |
| Breast cancer | MT-1/2 | doxorubicin | (Kepinska et al., 2018) |
| HCC | MT-1G, MT-1B, MT-1E, MT-1L, MT-1M | sorafenib | (Houessinon et al., 2016; Reeves, 2016; Sun et al., 2016) |
| | Total MTs | carboplatin | (Choi et al., 2004; Endo et al., 2004) |
| Lymphoma | MT-2A | gallium nitrate | (Yang and Chitambar, 2008; Yang et al., 2007) |
| Gastrointestinal Stromal Tumor | Total MTs | imatinib | (Perez-Gutierrez et al., 2007) |
| Cardioprotection* | Total MTs | doxorubicin | (Guo et al., 2014; Jing et al., 2011; Sun et al., 2001; Wang and Kang, 1999) |

1 *Not a cancer type but another biological functionality described for MTs.

2 *4.1. MTs and chemoresistance to platinum-based drugs*

3 To understand pretarget resistance in metallo-chemotherapeutics, it is necessary to investigate
4 the mechanisms responsible for metal–protein binding reactions. Platinum drugs, including
5 cisplatin, carboplatin, and oxaliplatin, are well-known chemotherapeutic drugs. They have
6 been utilized for the treatment of numerous human cancers, including brain, head and neck,
7 lung, breast, neuroblastoma, ovarian, bladder, and testicular cancers, for decades (Chin et al.,
8 1993; Esteban-Fernandez et al., 2008; Ravera et al., 2019; Wang et al., 2019). They are also
9 effective against various types of cancers, including carcinomas, germ cell tumors,
10 lymphomas, and sarcomas. Their mode of action has been linked to their ability to crosslink
11 the N₇ reactive center of purine residues and thereby cause DNA damage in cancer cells,
12 blocking cell division and subsequently inducing apoptosis (Dasari and Tchounwou, 2014).
13 The intracellular level of MTs may play an important role in regulating cellular
14 responsiveness to DNA-targeted antineoplastic agents (Basu, 2018). The antitumor activity of

1 cisplatin is believed to be due to its interaction with chromosomal DNA (Florea and
2 Büsselberg, 2011; Rebillard et al., 2008; Rocha et al., 2018). It is still uncertain which MT
3 isoforms are increased in cells with acquired resistance to platinum drugs. However, we show
4 in this review that MT levels correlate with the sensitivity of human tumors and cell lines to
5 platinum drugs. Cisplatin resistance remains a major impediment to the effective treatment of
6 many types of cancers. The cellular inactivation of cisplatin and subsequent sequestration can
7 be mediated by MTs that chelate platinum and prevent interaction with tumor cell DNA (Hou
8 et al., 2017; Maleckaite et al., 2019; Skowron et al., 2018; Wong and Stillman, 2018). In
9 1997, Hishikawa et al. suggested that MT expression in squamous cell carcinoma of the
10 esophagus is a major determinant of cisplatin resistance and may be a predictor of poor
11 prognosis (Hishikawa et al., 1997). To date, MT overexpression has been implicated in
12 cisplatin resistance in several types of cancer. MT-3 was initially thought to be unresponsive
13 to platinum drugs. However, we have recently shown a significant increase in
14 chemoresistance to cisplatin due to MT-3 upregulation in neuroblastoma (Rodrigo et al.,
15 2018). MT overexpression also predicts poor survival in bladder cancer patients. In patients
16 treated with cisplatin-based chemotherapy, survival was significantly poorer when tumors
17 expressed MT (Wulfing et al., 2007). Gumulec et al. reported that cisplatin-resistant prostate
18 cancer cells displayed a significant increase in MT expression and decreased p53 and Bax
19 (Gumulec et al., 2014a). These results, along with those of another study, may explain the
20 events leading to the development of cisplatin resistance in prostate cancer (Gumulec et al.,
21 2014a; Pekarik et al., 2013). Moreover, *MT-1G* was identified, *via* cDNA microarray, as a
22 candidate cisplatin-resistance-related gene for gastric cancer (Suganuma et al., 2003). In
23 NSCLC cells, *MT-1H* overexpression was shown to promote cisplatin resistance by
24 decreasing the induction of apoptosis (Hou et al., 2009). Interestingly, an analysis found
25 significantly higher serum MT levels in 25 patients with testicular cancer than in healthy

1 volunteers, and furthermore, a significant amount of platinum was bound to proteins in the
2 fraction of MT elution (Tariba et al., 2015).

3 The treatment of prostate cancer cells with Zn^{++} was found to increase MT expression, which
4 is significantly associated with resistance to cisplatin chemotherapy (Smith et al., 2006).

5 Other studies have shown that the elevation of MTs may be one mechanism of cisplatin
6 resistance in ovarian carcinoma (Cheng et al., 2006; Surowiak et al., 2005; Woolston et al.,
7 2010). Elevated MT expression in ovarian cancers treated with cisplatin-based regimens was
8 reported as an unfavorable prognostic factor for this treatment regimen (Surowiak et al.,
9 2007). MTs were also found to be stably expressed at increased levels in cisplatin-resistant
10 ovarian cancer cell lines compared with their cisplatin-sensitive counterparts (Kawahara et al.,
11 2019).

12 The repeated administration of cisplatin as a treatment for human bladder tumors is known to
13 exert lethal and renal toxicities (Li et al., 2016b). Bismuth pretreatment effectively prevented
14 cisplatin toxicity without affecting its antitumor activity against human bladder tumors. MT
15 levels induced by increasing the dose of bismuth in the kidneys maintained their substantially
16 high levels during the treatment (Kondo et al., 1992). These data strongly suggest a promising
17 protocol for chemotherapy using cisplatin with bismuth-based compounds – tissue-specific
18 MT inducers that display very low untoward toxicity – against advanced bladder cancer. In
19 addition, Chang et al. showed that bismuth Zn^{++} citrate potentially reduced cisplatin-induced
20 toxicity without compromising the anticancer effect through enhanced expression of MTs
21 (Chan et al., 2019).

22 Oxaliplatin and carboplatin derivatives of cisplatin have a similar mechanism of action but
23 differ in terms of structure and toxicity. Previously, it has been observed that the intracellular
24 mechanisms by which cells become resistant to carboplatin involve increased drug
25 detoxification by the thiol groups of MTs and improved tolerance to nuclear damage, leading

1 to a concomitant reduction in apoptosis and reduced accumulation of intracellular carboplatin
2 (Kukacka et al., 2008; Wheate et al., 2010). The examination of MT expression in tissue
3 biopsy specimens from HCC patients was shown to be useful in predicting the therapeutic
4 effect of carboplatin (Endo et al., 2004).

5 Oxaliplatin, similar to other platinum-based compounds, exerts its cytotoxic effect mostly
6 through DNA damage (Cao et al., 2020; Durmus et al., 2016; Ferreira et al., 2016; Hosseini et
7 al., 2019; Leonetti et al., 2019b; Mokady and Meiri, 2015; Wijdeven et al., 2016). Cancer cell
8 apoptosis is induced by the formation of DNA lesions, arrest of DNA replication, inhibition of
9 transcription, and triggering of immunologic reactions (Alcindor and Beauger, 2011). Using
10 novel strategies, Arriaga et al. showed that MT induction and Zn administration were feasible
11 to sensitize CRC cells to oxaliplatin (Arriaga et al., 2014). Wong et al. demonstrated the
12 ability of *MT-1A* to counteract transition metal complexes with cisplatin, emphasizing the
13 detrimental role of MTs as a major player in the reduced effectiveness of metal-based drugs
14 (Wong and Stillman, 2018).

15 Heptaplatin is a new platinum derivative gaining interest for its anticancer activity against
16 cisplatin-resistant cancer cell lines (Xu and Wang, 2016). Interestingly, gastric cancer cell
17 lines express different basal MT mRNA levels (Soo et al., 2011). Moreover, DNA
18 hypomethylation was proposed to be responsible for the higher basal *MT-2* mRNA levels in
19 the cisplatin-resistant tumor cell lines. Choi et al. showed reduced cytotoxicity of cisplatin and
20 carboplatin but not heptaplatin following MT induction with Zn. Heptaplatin was more
21 efficient than both cisplatin and carboplatin against cisplatin-resistant gastric cancer cell lines
22 and MT-overexpressing cell lines (Choi et al., 2004). Despite the absence of correlations
23 between MT overexpression and platinum-drug resistance in some types of malignant cancers
24 (Gansukh et al., 2013; Tuzel et al., 2015), most studies corroborated that elevated levels of
25 MTs may be a plausible mechanism of cisplatin resistance in cancer.

1 4.2. MTs and chemoresistance to anthracyclines

2 Anthracyclines are anticancer drugs that were originally derived from *Streptomyces* bacteria.
3 Their antitumor activity was established in the 1960s. The four most common anthracyclines
4 are Dox, daunorubicin, epirubicin and idarubicin (Marinello et al., 2018).

5 Dox is the lead compound of the anthracycline family. Cardiotoxicity limits anthracycline
6 dosing, and despite improved cancer patient outcomes, the cancer survivors are subject to
7 increased cardiovascular morbidity and mortality. The basic mechanisms of cardiotoxicity
8 may involve direct pathways for ROS generation and topoisomerase II inhibition as well as
9 other indirect pathways (McGowan et al., 2017). In this respect, it has been previously shown
10 that cardiomyocyte-targeted deletion of the *TOP2B* gene protected cardiomyocytes from Dox-
11 mediated double-strand DNA breaks and transcriptome alterations, both of which underlie
12 defective mitochondrial biogenesis and ROS generation (Zhang et al., 2012). Moreover,
13 cardiomyocyte-targeted deletion of *TOP2B* protected mice from Dox-induced progressive
14 heart failure, indicating that Dox-induced cardiotoxicity is mediated *via* topoisomerase-II β in
15 cardiomyocytes.

16 Studies using transgenic mice with high levels of antioxidants such as MTs, specifically in the
17 heart, have demonstrated that elevated cardiac antioxidant defense leads to decreased
18 anthracycline cardiotoxicity. Positive correlations between histopathological lesions,
19 apoptosis and MT expression were observed by Chmielewska et al. (Chmielewska et al.,
20 2015). The results suggested that MT expression had protective and anti-apoptotic effects in
21 renal proximal tubular cells under Dox treatment. MT-dependent protection against
22 anthracycline-induced cardiotoxicity is related to its anti-apoptotic effects achieved by
23 inhibiting both p38-MAPK-mediated and mitochondrial cytochrome c release-mediated
24 apoptotic signaling (Kang, 2007). *MT-2* downregulation in MCF-7 cells resulted in increased
25 chemosensitivity of these cells to Dox (Kepinska et al., 2018). Another study has shown that

1 cardiac-specific MT-overexpressing transgenic mice are highly resistant to acute
2 cardiotoxicity induced by Dox (Sun et al., 2001). In addition, MT-3 overexpression attenuated
3 the effect of Dox on cell proliferation pathways in metastatic prostate cancer cell lines (Juang
4 et al., 2013). The reduced cytotoxic effect of Dox in MTs overexpressing cardiomyocytes was
5 correlated with the inhibition of lipid peroxidation induced by the drug (Wang and Kang,
6 1999). MT overexpression protected against Dox-induced inhibition of PGC-1 α , a key
7 regulator of mitochondrial biogenesis, and its downstream factors, including mitochondrial
8 transcription factor A (Guo et al., 2014). Heger et al. showed in an exhaustive bibliographic
9 review that MTs, as scavengers of ROS, regulated anthracycline chemoresistance in cancer
10 and can also be used as a new cardioprotective therapeutic agent (Heger et al., 2016). In
11 conclusion, elevated MT levels confer chemoresistance to anthracycline cytotoxicity through
12 a mechanism involving the anti-apoptotic action of MTs. The MT redox cycle and Zn⁺⁺
13 homeostasis most likely constitute the MT-involved antioxidant defense.

14 *4.3. MTs and resistance to tyrosine kinase inhibitors (TKIs)*

15 TKIs are a class of chemotherapeutic agents that inhibit or block one or more of the protein
16 tyrosine kinases. TKIs are a family of small molecules or peptides with the ability to inhibit
17 either cytosolic or receptor tyrosine kinases (Leonetti et al., 2019a; Roskoski, 2020). Many
18 TKIs have been developed and approved across a wide range of tumor types to determine the
19 critical roles of tyrosine kinases in regulating cellular signaling and tumor growth in patients
20 (Gillis and McLeod, 2016). Inhibition by this class of cytotoxic agents is mediated through
21 direct competition for ATP binding in the tyrosine kinase domain (genistein, lavendustin,
22 imatinib, erlotinib, gefitinib, sorafenib), allosteric inhibition of tyrosine kinases (lavendustin
23 A), inhibition of ligand binding to receptor tyrosine kinases (ecetuximab), inhibition of
24 tyrosine kinase interaction with other proteins, or destabilization of the tyrosine kinase

1 (herbimycin A and radicicol) (Reeves, 2016). However, acquired resistance of TKIs to
2 targeted therapies inevitably occurs (Camidge et al., 2014).

3 Sorafenib was originally identified as an inhibitor of multiple oncogenic kinases and remains
4 the only approved systemic therapy for advanced HCC. The antitumor efficiency of sorafenib
5 correlates with the inhibition of the Ser/Thr kinase Raf and several receptor tyrosine kinases,
6 including VEGFR and EGFR (Siegel et al., 2010). Currently, there is a lack of articles
7 regarding the MT mechanism in sorafenib-resistant cancers. However, two studies
8 demonstrated that *MT-1G* and *MT-1H* isoforms act as tumor suppressors in HCC development
9 (Zeng et al., 2018; Zheng et al., 2017). Sun et al. demonstrated that upregulated *MT-1G*
10 expression protects HCC cells from sorafenib and facilitates cancer progression by inhibiting
11 lipid peroxidation-mediated ferroptosis. Thus, the modulation of *MT-1G* expression is a
12 potential therapeutic strategy to overcome acquired resistance to sorafenib in cancer (Sun et
13 al., 2016). Genes of the *MT-1* family are also induced in the HCC cell line Huh7 upon
14 exposure to sorafenib. Houessinon et al. examined the clinical relevance of characterizing the
15 regulation of *MT-1G* in five tumor explants prepared from surgical HCC samples. The protein
16 levels of MT-1 increased in the serum of HCC patients receiving sorafenib (Houessinon et al.,
17 2016). The mRNA expression and protein expression of *MT-1G* are both markedly induced
18 by sorafenib but not by other clinically relevant protein kinase inhibitors (*e.g.*, erlotinib,
19 gefitinib, tivantinib, vemurafenib, selumetinib, imatinib, masitinib, and ponatin). However,
20 Pérez-Gutiérrez et al. showed that the differences in P-glycoprotein and MT expression could
21 help to explain the observed response to systemic imatinib chemotherapy in gastrointestinal
22 stromal tumors and leiomyosarcomas (Perez-Gutierrez et al., 2007).

23 *4.4. MTs and resistance to mitomycin C*

24 Mitomycin C, a potent DNA cross-linker, is one of the most commonly used agents in bladder
25 cancer and has limited side effects. Specifically, intravesical instillation of mitomycin C

1 following a transurethral resection of a bladder tumor constitutes a standard treatment
2 modality in the management of superficial transitional cell carcinoma of the urinary bladder.
3 However, MT overexpression predicts the resistance of superficial transitional cell carcinoma
4 of the bladder to intravesical mitomycin C therapy (Lynn et al., 2003). Another study
5 suggested that cross-resistance to cisplatin and carboplatin in a mitomycin C-resistant human
6 bladder cancer cell line may be due to the overexpression of MT-2-encoding mRNA (Singh et
7 al., 1995).

8 *4.5. MTs and resistance to other anticancer drugs*

9 Gemcitabine is a chemotherapeutic drug used for the treatment of NSCLC. MTs were highly
10 associated with the sensitivity of gemcitabine in NSCLC (Chunhong et al., 2018).
11 Overexpression of the MT-1G isoform sensitized CRC cell lines to the chemotherapeutic
12 agent 5-fluorouracil in combination with oxaliplatin, in part through enhancing p53 and
13 repressing NF- κ B activity (Arriaga et al., 2014).

14 Irinotecan, a camptothecin derivative, is a DNA topoisomerase I inhibitor that is active
15 against gastric cancer tumors (Chun et al., 2004b). Chun et al. suggested that irinotecan-
16 induced upregulation of *MT-IX* might be associated with irinotecan resistance in patients with
17 gastric cancer (Chun et al., 2004a).

18 Several clinical trials have shown gallium nitrate to be an active agent in the treatment of
19 lymphoma. MT expression contributed to the development of gallium drug resistance.
20 Gallium nitrate induced MT-2 and ZnT-1 expression in lymphoma cells. A role for MTs in
21 modulating the antineoplastic activity of gallium was confirmed by showing that the induction
22 of MT expression by Zn⁺⁺ provided partial protection against the cytotoxicity of gallium and
23 that the level of endogenous MTs in lymphoma cell lines correlated with their sensitivity to
24 gallium nitrate (Yang and Chitambar, 2008; Yang et al., 2007).

25 **5. Conclusions and future perspectives**

1 Cancer remains a major cause of mortality worldwide. Currently, drug resistance in cancer is
2 the foremost threat to curative therapeutics. MTs contribute to the development of drug
3 resistance through a variety of mechanisms in many types of cancers. MTs are proteins with
4 an inexhaustible spectrum of biological functions. Despite their importance for the physiology
5 of healthy cells, MTs also play a substantial role in various aspects of human malignancies.
6 These proteins have functions such as metal ion homeostasis and detoxification, antioxidation
7 against reactive oxygen species, protection against DNA damage, and the regulation of cell
8 growth, proliferation, angiogenesis, and apoptosis. This review article shows that many
9 independent groups of investigators found direct and indirect correlations between MTs and
10 chemoresistance.

11 In addition to ROS detoxification, MT–drug interactions can be mediated directly in the case
12 of metals (*e.g.*, platinum in cisplatin and gallium in gallium nitrate) or mediated by Zn
13 homeostasis (*e.g.*, by controlling p53 DNA-binding domain folding) or by neutralizing
14 reactive drug intermediates *via* MT sulfhydryl oxidation (*e.g.*, the semiquinone form of Dox
15 and the mitosene form of mitomycin C). Tyrosine kinase inhibitors serve as an alternative and
16 indirect mechanism of resistance through MT expression modulation.

17 A great deal of research remains to be performed to fully unravel the molecular roles of MTs
18 in this aspect. These future studies should focus on elucidating the distinct functions of
19 individual MT isoforms, which could be useful not only for improved diagnostics but also as
20 *bona fide* druggable targets for precision cancer therapy (or for combination therapy). Some
21 insights were also noted regarding the usefulness of combining alternative compounds (*e.g.*,
22 bismuth-based compounds with cisplatin) for modulating MT activity, thus reducing drug
23 toxicity without compromising anticancer effects. In addition, exploration of the regulatory
24 mechanisms responsible for the expression of MT isoforms might provide substantial insights
25 into their importance in cancer biology and therapeutics. In this review, we have shown MTs

1 as molecular players that change sides once carcinogenesis occurs. The identification of that
2 upstream molecular switch is surely connected with the transcriptional regulation of MT
3 isoforms, and it is possible that various scenarios can arise for different types of cancer. In-
4 depth knowledge on the regulation of MTs may bring hope for overcoming chemoresistance
5 in cancer.

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13 **Declaration of Competing Interests**

14 The authors declare no conflicts of interest.

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